

# HIV-Associated Renal Diseases and Highly Active Antiretroviral Therapy–Induced Nephropathy

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Renal disease is becoming an increasingly prevalent entity in human immunodeficiency virus (HIV)–infected patients; it occurs in a variety of clinical settings and is associated with histopathological changes. HIV-related renal impairment can present as acute or chronic kidney disease; it can be caused directly or indirectly by HIV and/or by drug-related effects that are directly nephrotoxic or lead to changes in renal function by inducing metabolic vacuolopathy and renal damage. Acute renal failure is frequently caused by the toxic effects of antiretroviral therapy or nephrotoxic antimicrobial substances used in the treatment of opportunistic infections. Chronic renal disease can be caused by multiple pathophysiological mechanisms, leading to HIV-associated nephropathy, a form of collapsing focal glomerulosclerosis, thrombotic microangiopathy, and various forms of immune complex glomerulonephritis. The increase in life expectancy and alteration of lipid metabolism due to receipt of highly active antiretroviral therapy are expected to result in an increased prevalence of diabetes and hypertension and, thus, to secondary diabetic and hypertensive renal damage. Antiretroviral agents, such as indinavir and tenofovir, have been associated with nephrotoxic drug effects that have been shown to be reversible in most cases. In this article, we review the current knowledge about acute and chronic HIV-associated renal disease, metabolic alterations and related nephropathies, and toxic drug effects of combination antiretroviral pharmacotherapy.

Nephropathy is a common finding in patients infected with HIV, and it necessitates increased surveillance and adaptation of dosages of HIV drugs. Direct effects of HIV seem to play a major role in the development of HIV-associated nephropathy (HIVAN) and thrombotic microangiopathy. Improved survival among patients with HIV infection is anticipated to result in an increase in the long-term development of HAART-associated metabolic complications, such as diabetes and dyslipidemia, which, in turn, can contribute to vascular changes and decreased renal function. Since the introduction of HAART, a variety of renal side effects and adverse drug reactions have been recognized and vary

from the development of proteinuria to acute renal failure.

## ACUTE RENAL FAILURE IN HIV-INFECTED PATIENTS

Acute renal failure in HIV-infected persons can be caused by the same mechanisms that cause it in HIV-uninfected patients. Data from the HAART era are rare, although a recent study that evaluated the incidence and etiology in a prospective analysis of 754 HIV-infected patients reported an incidence of 5.9 cases of acute renal failure per 100 patient-years [1]. Drugs used for treatment of HIV infection that are associated with nephrotoxicity include aminoglycosides, amphotericin, foscarnet, trimethoprim-sulfamethoxazole, tenofovir, indinavir, and acyclovir [2]. Furthermore, acute renal failure may be related to thrombotic thrombocytopenic purpura–hemolytic uremic syndrome or pharmacotherapy, which will be discussed below with the respective substances. Acute renal failure is a common

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finding in HIV-infected patients and is associated with advanced stages of HIV infection (i.e., CD4 cell count of  $<200$  cells/mm<sup>3</sup> and HIV RNA level of  $>10,000$  copies/mL), hepatitis C virus coinfection, and a history of antiretroviral treatment [1].

## CHRONIC RENAL DISEASE IN HIV-INFECTED PATIENTS

The prevalence of chronic kidney disease in the various stages of HIV infection is difficult to assess. Proteinuria and elevated creatinine level have been found in 7.2% [3] to 32% [4] of HIV-seropositive patients and were associated with an increased rate of death in a study of 2038 female HIV-infected patients [5]. Proteinuria still remains a nonspecific finding in HIV-infected patients. Autopsy studies yield a prevalence of up to 43% of pathological changes on histological examination [6]. Both autopsy and biopsy studies may be limited by bias of selecting subjects presenting with apparent renal disease. HAART has been found to reduce progression from AIDS to end-stage renal disease in patients of African descent by 38%, yet a significant increase in the prevalence of end-stage renal disease (ESRD) associated with an increase in the prevalence of HIV infection in this population has been predicted [7]. Even though exact epidemiologic data are missing because of the use of different screening techniques, chronic kidney disease in HIV-infected patients is a common and clinically relevant finding. The cause of chronic renal disease in HIV-infected patients can be difficult to assess on clinical grounds alone and can most often only be determined by renal biopsy.

## NEPHROPATHY ASSOCIATED WITH HIV INFECTION

Several types of renal disease seem to be directly or indirectly caused by HIV: classic HIVAN, HIV-associated thrombotic microangiopathy, and HIV-associated, immune-mediated glomerulonephritides [2].

**HIVAN.** Classic HIVAN is a syndrome caused by focal sclerosing glomerulopathy with severe proteinuria, renal failure, and rapid progression to ESRD. It has become the most common cause of ESRD in HIV-1-seropositive patients. HIVAN primarily occurs in patients of African descent [8–11], suggesting a genetic predisposition to the disease. Duffy antigen/receptor for chemokines has controversially been discussed as a candidate gene involved in the development of HIVAN [12, 13]. The estimated prevalence of HIVAN has ranged from 3.5% in clinical studies to 12% in autopsy studies [14]. Because HIVAN typically occurs late in the course of HIV-1 infection [15], risk factors for the development of HIVAN include a CD4 cell count  $<200$  cells/mm<sup>3</sup> and a high viral burden.

Clinical features of this syndrome include advanced renal failure and proteinuria (the protein level is often—but not

necessarily—at a nephrotic level [ $>3$  g/day]) [16, 17], a lack of peripheral edema despite the severe loss of protein, and, frequently, enlarged kidneys visible on renal ultrasound [9].

Renal biopsy is the only means to establish the diagnosis of HIVAN. Characteristic histological findings include collapsing focal and segmental glomerulosclerosis, tubular epithelial atrophy with microcystic dilatation of the tubules [18], and lymphocytic interstitial infiltration. Viral infection of renal cells seems to play an important role in the pathogenesis of HIVAN. In 1989, Cohen et al. [19] reported detection of HIV-1 in renal epithelial cells by DNA in situ hybridization. Transgenic mouse models have been described in which expression of a *gag/pol*-deleted provirus leads to HIVAN-like histological changes [20] or in which the single HIV-1 gene *nef* can lead to immunodeficiency and renal disease [21]. HIV can be detected in biopsy specimens from the renal epithelium in humans and can replicate in renal epithelial cells, even in patients who are receiving effective antiretroviral therapy [22]. In 2002, a study was published demonstrating a tissue-specific evolution of HIV-1 gp120 DNA sequences in renal epithelial tissues that was different from those observed in PBMCs, suggesting that renal epithelial cells are capable of complete viral replication and represent a separate compartment for HIV-1 [23].

Without adequate treatment, the prognosis of HIVAN is poor. Usually, HIV-associated nephropathy is diagnosed at a late stage, and untreated patients frequently have progression to ESRD within a few months [10]. The most effective therapeutic option seems to be HAART. Cases have been documented in which initiation of antiretroviral treatment led to clinical and histological remission within a few weeks [24]; other patients ended dialysis treatment [25]. HAART also seems to have a protective renal effect. In a 12-year follow-up study of 3976 HIV-1-seropositive patients, a 60% reduction in the risk of development of HIVAN associated with HAART was found [26]. Although there is strong observational data supporting a role for HAART in the treatment of HIVAN, no prospective, randomized, controlled trials have been performed that support a beneficial effect of HAART or of other medical therapies. Several studies with limitations resulting from lack of randomization or small size seem to support a beneficial effect of angiotensin-converting enzyme inhibitors [27]. Because trials involving prednisone have been rare in the HAART era, there is not enough evidence to generally support treatment of HIVAN with corticosteroids [28].

**Thrombotic microangiopathy.** Thrombotic microangiopathy, hemolytic uremic syndrome, and thrombotic thrombocytopenic purpura present a spectrum of diseases characterized by hemolytic anemia, thrombocytopenia, and renal insufficiency and clinical features, such as fever and neurological manifestations. Several reports have linked thrombotic microangiopathy

to HIV infection, suggesting that HIV proteins may mediate endothelial dysfunction, leading to platelet deposition in the microvasculature [29]. Several potential underlying mechanisms have been discussed: HIV-1 p24 antigen has been detected in endothelial cells in patients with thrombotic thrombocytopenic purpura [30]; secretion of inflammatory cytokines, such as TNF- $\alpha$  and IL-1 $\beta$  [31], leading to endothelial cell retraction has been documented; and renal endothelial cell apoptosis and inhibition of von Willebrand factor–cleaving protease have been linked to HIV-induced thrombotic microangiopathy [32].

Typical clinical features of thrombotic thrombocytopenic purpura–hemolytic uremic syndrome are onset at ~35 years of age, a predominance among male subjects, and poorer outcome, compared with idiopathic forms of the disease [33]. Therapeutic options consist of plasma infusion and plasmapheresis, which have had variable success. Other attempted therapies include glucocorticoids, immunoglobulin infusions, antiplatelet drugs, vincristin, and splenectomy, although general treatment recommendations are lacking.

**Immune complex–mediated glomerulonephritis.** A multitude of immune complex–mediated glomerulonephritides have been reported as causes of chronic kidney disease in HIV-infected patients. The prevalence of HIV-associated, immune complex–mediated glomerulonephritides has been estimated to be 15%–80%. A study of 60 biopsy specimens found that some form of immune complex–mediated glomerulonephritis was present in 37% of biopsy specimens, sometimes concordant with HIVAN-like changes. The authors classified their findings into 4 categories: immune complex–mediated glomerulonephritis, IgA nephritis, mixed sclerotic/inflammatory disease, and lupus-like syndrome [34]. Immune complex–mediated glomerulonephritis may present as postinfectious glomerulonephritis, membranous nephritis, IgA nephritis, fibrillary glomerulonephritis, immunotactoid glomerulopathy, and membranoproliferative glomerulonephritis [2].

IgA nephritis seems to be more prevalent among European patients, with mesangial IgA deposits being detected in 7.75% of all HIV-infected patients in a postmortem study [35]. A recent report estimated that the prevalence of lupus-like nephritis, characterized by immunoglobulin (IgG, IgA, and IgM) and complement (C3 and C1q) deposits, in the absence of serologic markers for systemic lupus erythematoses, was 17% [36].

In general, HIVAN is mainly limited to patients of African descent, whereas most cases of renal disease in the white population seem to be immune complex–mediated glomerulonephritis [37]. Despite the lack of randomized trials, patients with HIV-associated, immune complex–mediated glomerulonephritis seem to benefit from treatment with angiotensin-converting enzyme inhibitors, glucocorticoids, and antiretrovirals [38, 39].

## **METABOLIC ALTERATIONS ASSOCIATED WITH ANTIRETROVIRAL TREATMENT**

Metabolic alterations associated with HAART may lead to significant elevations in serum lipid levels, accelerating the development of diabetes and the metabolic syndrome. Long-term metabolic changes will possibly cause an increase in diabetic and hypertensive renal disease as well as vascular complications.

**Diabetes mellitus.** Antiretroviral therapy has been associated with impaired glucose tolerance [40, 41]. A study of 17,852 HIV-seropositive patients evaluated the prevalence of cardiovascular risk factors and reported a prevalence of diabetes of 2.5% that was significantly associated with antiretroviral treatment [42]. A controlled study evaluated 1689 patients and reported an incidence of diabetes of 14% among patients who were receiving antiretroviral therapy and demonstrated a 4-fold risk of developing diabetes in the HIV-infected group [43]. The apparent increase in the incidence of diabetes may be the result of methodological differences or may be an expression of the elevated risk of developing diabetes associated with HAART-induced metabolic changes and an increased survival duration. Data on the prevalence of secondary damage due to diabetes, such as nephropathy, retinopathy, or polyneuropathy, are lacking.

**Hypertension.** The findings of studies assessing HAART-associated hypertension are inconsistent [42]. A recent study that observed a cohort of 5578 patients during 1984–2003 revealed an incidence of hypertension in 7.3% among HIV-seropositive patients who had a systolic blood pressure >140 mm Hg, and the incidence began to increase significantly after 2 years of antiretroviral therapy. The risk of developing hypertension was maximally elevated (OR, 1.7) after >5 years of treatment with HAART, compared with the risk among HIV-infected patients who did not receive treatment (OR, 0.79) [44].

A study that evaluated 98 renal biopsy specimens obtained from HIV-1–seropositive patients with clinical evidence of nephropathy revealed diabetic glomerulopathy in 6 patients (6.1%) and hypertensive nephrosclerosis in 4 patients (4.1%) [45]. Additional biopsy studies can only be performed ethically in the stage of severe renal impairment, thus possibly leading to false conclusions resulting from selection bias. Because moderate forms of diabetic and hypertensive glomerulopathy cannot be confirmed by examination of biopsy specimens, the true incidence of metabolic renal disease remains unknown.

## **RENAL ADVERSE EFFECTS OF HAART**

Renal damage caused by antiretroviral drugs can result in a variety of toxic drug effects presenting as acute renal failure, tubular necrosis, kidney stones, or chronic renal disease.

## Protease Inhibitors

**Indinavir.** Indinavir is the protease inhibitor that has been most frequently associated with adverse renal effects, including nephrolithiasis, crystalluria, dysuria, papillary necrosis, and acute renal failure [46]. In 1997, a study demonstrated that urinary crystals composed of indinavir occurred in 20% of all patients who used indinavir, and the condition progressed to nephrolithiasis in 3%. Other patients without detectable stone formation (12.5%) presented with symptoms of dysuria and flank pain [47]. A study of 1219 patients estimated that the incidence of urological complications was 8.3 cases per 100 treatment-years [48]. Stone formation can occur in any structure of the kidney and the urinary tract, with risk factors being a urine pH >6, low lean body mass level, high concentrations of indinavir, environmental influences (such as a warm climate) [48], concomitant treatment with trimethoprim-sulfamethoxazole [49] or acyclovir [50], and concordant chronic hepatitis B or C virus infection [51]. Antiretroviral therapy given in combination with low-dose ritonavir seems to increase the renal toxicity of indinavir [52]. The safest way to avoid urological symptoms seems to be maintenance of a high urinary output of 1500 mL/day. After acute renal failure occurs, it appears to be safe to restart indinavir treatment once adequate rehydration is established. Acidification of the urine is expected to be poorly tolerated and possibly harmful [53]; thus, no recommendation to lower the urinary pH has been issued to date.

An increase in the serum creatinine level frequently preceded by sterile leukocyturia has been observed with indinavir treatment in 14%–33% of patients [54–58]. Most urologic symptoms and elevations in serum creatinine levels normalize within weeks after the discontinuation of indinavir, although irreversible renal toxicity has been reported. Interstitial nephritis was demonstrated in renal biopsy specimens; this can have a self-limited course [59, 60] or lead to interstitial fibrosis [61] and renal atrophy [62].

**Ritonavir.** Case reports have linked ritonavir use to reversible renal failure [63, 64]. All reported patients who showed an increase in the serum creatinine level received ritonavir at a therapeutic dose of 800–1200 mg per day. In 1 patient who had acute renal failure after exposure to ritonavir, rechallenge with a lower dose of ritonavir (200 mg) led to a second (yet reversible) decline in renal function [65]. The majority of reported patients had received concomitant medication with potentially nephrotoxic drugs or had other underlying renal pathology, such that specific cases of nephrotoxicity attributable to ritonavir use have not been definitely established.

**Saquinavir and nelfinavir.** Saquinavir and nelfinavir have been demonstrated to have a generally safe renal safety profile in controlled trials, but for both drugs, there have been single case reports suggesting the potential for inducing renal calculi

[66]. Analysis of a urinary stone revealed a composition of 99% nelfinavir and 1% indinavir—a drug that had been discontinued before treatment with nelfinavir had been started [67].

**Atazanavir.** Atazanavir has not been associated with renal toxicity in clinical trials. However, a recent case report described a patient with interstitial nephritis and reversible acute renal failure after addition of atazanavir to an otherwise stable HAART regimen. The therapeutic regimen included potentially nephrotoxic drugs, and examination of a renal biopsy specimen revealed histological changes of HIVAN, with collapsing glomerulosclerosis and interstitial nephritis [68].

**Other protease inhibitors.** Protease inhibitors, such as amprenavir, fosamprenavir, and lopinavir, have not been associated with severe renal disease to date.

## Nucleotide Reverse-Transcriptase Inhibitors

Tenofovir, adefovir, and cidofovir are acyclic nucleoside phosphonates that have been associated with renal tubular damage. The renal adverse effects may cause a variety of clinical presentations varying from tubule cell death, such as acute tubular necrosis, to possibly reversible tubular dysfunction, such as Fanconi syndrome.

A number of case reports linked tenofovir to tubular toxicity and Fanconi syndrome with variable severity [69–80]. Fanconi syndrome consists of a generalized defect of membrane transporters in the proximal tubule, leading to renal loss of glucose (despite a normal serum glucose concentration), as well as a loss of phosphate, calcium, uric acid, amino acids, bicarbonates, and tubular proteins [81]. Most cases associated with tenofovir use do not meet the criteria for Fanconi syndrome but present as severe tubular dysfunction with an elevation in creatinine levels, hypophosphatemia, and glycosuria [79]. Renal biopsy reveals tubular damage or acute tubular necrosis without involvement of the glomeruli [69–72]. Risk factors include low body weight, impaired renal function at baseline, and concomitant receipt of nephrotoxic drugs [77, 78]. In general, tubular dysfunction is reversible after withdrawal of tenofovir [78, 79], although persistent renal damage with impairment of renal function has been reported [17, 72]. Approximately 55 cases of tubular dysfunction in association with tenofovir use have been reported in the literature. The incidence of renal tubular dysfunction is difficult to assess, because these cases are anecdotal, and the incidence has not been reported in larger, well-controlled studies.

Recent reports have linked HAART regimens that contain tenofovir to a mild, time-dependant elevation in the serum creatinine level [82–84] and a decrease in the glomerular filtration rate. One study of 174 patients found a lower mean glomerular filtration rate, as calculated by creatinine clearance (97 mL/min [1.73 m<sup>2</sup>] vs. 107 mL/min [1.73 m<sup>2</sup>]), with 38%

of patients showing an impaired glomerular filtration rate in the tenofovir arm compared to 29% in the control group [85]. The decrease in glomerular filtration rate in those on a tenofovir-containing regimen remained well within the normal range of glomerular filtration rate and did not lead to discontinuation of tenofovir.

Nephrotoxicity may be caused by an imbalance of influx and efflux of drugs into renal cells, thus leading to an increase in the intracellular drug concentration [86]. Influx of tenofovir into renal tubular cells is mediated via the human organic anion transporter 1 [87]. Efflux pumps responsible for the apical efflux of tenofovir out of proximal tubule cells have not been identified, but on the basis of its transportation of adefovir [88], it has been hypothesized that the multidrug-resistance protein 4 is responsible for transporting tenofovir into the tubular lumen [89]. Because ritonavir has been associated with inhibition of multidrug-resistance protein 2, an efflux pump for organic anions, it has been hypothesized that the interactions between ritonavir and multidrug-resistance proteins may result in a decreased efflux of tenofovir, leading to a toxic increase in intracellular drug concentration [89]. This hypothesis is supported by clinical reports describing tenofovir-associated tubular renal dysfunction mainly in patients receiving salvage therapy who are receiving concurrent ritonavir treatment [89]. Inhibition of multidrug-resistance protein 4 by ritonavir has not yet been described [90, 91].

In a 3-year study that compared 602 therapy-naive patients with a backbone of lamivudine and efavirenz treatment with either tenofovir or stavudine, no difference in the incidence of renal dysfunction was found [92]. A recent evaluation of the same study population demonstrated that only 2 patients in the tenofovir arm developed grade 2 nephrotoxicity (defined as increase in the serum creatinine level of 2.1–3.0 mg/dL) that resolved with continued treatment. Overall results were similar to those for the control group, and severe nephrotoxicity did not occur [79]. In all randomized, double-blinded studies, tenofovir has been demonstrated to have a renal safety profile similar to that of other combination therapies [93–95] and to have an overall low potential for nephrotoxicity. Renal tubular dysfunction seems to be an uncommon but important adverse effect of therapy with tenofovir.

### **Nucleoside Reverse-Transcriptase Inhibitors**

Renal toxicity associated with the use of nucleoside analogues is generally rare. Case reports have demonstrated that didanosine [96, 97] and lamivudine-stavudine treatment have been associated with tubular dysfunction or Fanconi-like syndrome [98]. Another report described acute renal failure and biopsy-proven interstitial nephritis after exposure to abacavir [99].

### **Nonnucleoside Reverse-Transcriptase Inhibitors**

Data regarding renal toxicity with nonnucleoside reverse-transcriptase inhibitors use are limited, because nevirapine, efavirenz, and delavirdine have been demonstrated to have a safe renal profile in controlled trials. A single case report linked efavirenz to renal toxicity on the basis of a hypersensitivity reaction involving pneumonitis, hepatitis, and interstitial nephritis; symptoms recurred after a rechallenge. [100].

### **Fusion Inhibitors**

Enfuvirtide has not been associated with severe renal adverse effects. In a safety analysis of 663 patients in the T-20 versus Optimized Regimen Only (TORO)-1 and TORO-2 trials, 1 case patient who had a history of diabetes, proteinuria, and hematuria developed membranoproliferative glomerulonephritis [101].

## **SUMMARY**

Renal pathology in HIV-infected persons can be caused by a variety of mechanisms leading to a broad spectrum of clinical disease. HIV itself seems to directly mediate the development of HIVAN and thrombocytopenic purpura. Other pathophysiological pathways comprise indirect viral effects, such as renal immune complex deposition. Long-term survival contributes to an increase in HAART-induced metabolic alterations, diabetes, and hypertension and is likely to be associated with an increase in secondary renal damage, such as hypertensive nephrosclerosis and diabetic glomerulopathy. For the majority of antiretroviral substances, HAART-related effects on renal function do not seem to be highly relevant with regard to nephrotoxicity. Still, indinavir has been associated with frequent renal and urological adverse effects and with stone formation. Despite a safe renal profile in clinical trials, a number of case studies have reported tubular dysfunction associated with tenofovir use. In large, controlled clinical trials, the incidence of renal abnormalities in the treatment arm did not differ from that in the control arm.

The expected increase in the incidence of renal disease and end-stage renal failure might, to some extent, be prevented by close monitoring of renal function, including an estimation of creatinine clearance before HAART is initiated, and continued monitoring during treatment—specifically, for patients receiving late-stage HIV therapy and in the context of a large number of coadministered medications. Any changes in serum creatinine level or the development of proteinuria should lead to an early investigation of the cause. Prompt recognition should induce studies that evaluate the different therapeutic options in larger clinical trials. Until then, clinicians should be aware of the increasing numbers of HIV-seropositive patients with renal pathology who frequently require special attention, as well as the need to alter doses for patients with renal impairment.

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## References

1. Franceschini N, Napravnik S, Eron JJ Jr, Szczech LA, Finn WF. Incidence and etiology of acute renal failure among ambulatory HIV-infected patients. *Kidney Int* **2005**;67:1526–31.
2. Kimmel PL, Barisoni L, Kopp JB. Pathogenesis and treatment of HIV-associated renal diseases: lessons from clinical and animal studies, molecular pathologic correlations, and genetic investigations. *Ann Int Med* **2003**;139:214–27.
3. Gardner LI, Holmberg SD, Williamson JM, et al. HIV Epidemiology Research Study Group: Development of proteinuria or elevated serum creatinine and mortality in HIV-infected women. *J Acquir Immune Defic Syndr* **2003**;32:203–9.
4. Szczech LA, Gange SJ, van der Horst C, et al. Predictors of proteinuria and renal failure among women with HIV infection. *Kidney Int* **2002**;61:195–202.
5. Szczech LA, Hoover DR, Feldman JG, et al. Association between renal disease and outcomes among HIV-infected women receiving or not receiving antiretroviral therapy. *Clin Infect Dis* **2004**;39:1199–206.
6. Hailemariam S, Walder M, Burger HR, et al. Renal pathology and premortem clinical presentation of Caucasian patients with AIDS: an autopsy study from the era prior to antiretroviral therapy. *Swiss Med Wkly* **2001**;131:412–7.
7. Schwartz EJ, Szczech LA, Ross MJ, Klotman ME, Winston JA, Klotman PE. Highly active antiretroviral therapy and the epidemic of HIV-end-stage renal disease. *J Am Soc Nephrol* **2005**;16:2412–20.
8. Bourgoignie JJ, Ortiz-Interian C, Green DF, Roth D. Race, a cofactor in HIV-1-associated nephropathy. *Transplant Proc* **1989**;21:3899–901.
9. Bourgoignie JJ, Meneses R, Ortiz C, Jaffe D, Pardo V. The clinical spectrum of renal disease associated with human immunodeficiency virus. *Am J Kidney Dis* **1988**;12:131–7.
10. Carbone L, D'Agati V, Cheng JT, Appel GB. Course and prognosis of human immunodeficiency virus-associated nephropathy. *Am J Med* **1989**;87:389–95.
11. Laradi A, Mallet A, Beaufils H, Allouache M, Martinez F. HIV-associated nephropathy: outcome and prognosis factors. Groupe d'Etudes Nephrologiques d'Ile de France. *J Am Soc Nephrol* **1998**;9:2327–35.
12. Liu XH, Hadley TJ, Xu L, Peiper SC, Ray PE. Up-regulation of Duffy antigen receptor expression in children with renal disease. *Kidney Int* **1999**;55:1491–500.
13. Woolley IJ, Kalayjian R, Valdez H, et al. HIV nephropathy and the Duffy antigen/receptor for Chemokines in African Americans. *J Nephrol* **2001**;14:384–7.
14. Ahuja TS, Borucki M, Funtanilla M, Shahinian V, Hollander M, Rajaraman S. Is the prevalence of HIV-associated nephropathy decreasing? *Am J Nephrol* **1999**;19:655–9.
15. Winston JA, Klotman ME, Klotman PR. HIV-associated nephropathy is a late, not early, manifestation of HIV-1 infection. *Kidney Int* **1999**;55:1036–40.
16. D'Agati V, Appel GB. HIV infection and the kidney. *J Am Soc Nephrol* **1997**;8:138–52.
17. Atta MG, Choi MJ, Longenecker JC, et al. Nephrotic range proteinuria and CD4 count as noninvasive indicators of HIV-associated nephropathy. *Am J Med* **2005**;118:1288.
18. Ross MJ, Bruggeman LA, Wilson PD, Klotman PE. Microcyst formation and HIV-1 gene expression occur in multiple nephron segments in HIV-associated nephropathy. *J Am Soc Nephrol* **2001**;12:2645–51.
19. Cohen AH, Sun NC, Shapshak P, Imagawa DT. Demonstration of human immunodeficiency virus in renal epithelium in HIV-associated nephropathy. *Mod Pathol* **1989**;2:125–8.
20. Dickie P, Felser J, Eckhaus M, et al. HIV-associated nephropathy in transgenic mice expressing HIV-1 genes. *Virology* **1991**;185:109–19.
21. Hanna Z, Kay DG, Rebai N, Guimond A, Jothy S, Jolicoeur P. Nef harbors a major determinant of pathogenicity for an AIDS-like disease induced by HIV-1 in transgenic mice. *Cell* **1998**;95:163–75.
22. Bruggeman LA, Ross MD, Tanji N, et al. Renal epithelium is a previously unrecognized site of HIV-1 infection. *J Am Soc Nephrol* **2000**;11:2079–87.
23. Marras D, Bruggeman LA, Gao F, et al. Replication and compartmentalization of HIV-1 in kidney epithelium of patients with HIV-associated nephropathy. *Nat Med* **2002**;8:522–6.
24. Kichner JT. Resolution of renal failure after initiation of HAART: 3 cases and a review of the literature. *AIDS Read* **2002**;12:103–5, 110–2.
25. Wali RK, Drachenberg CI, Papadimitriou JC, Keay S, Ramos E. HIV-1-associated nephropathy and response to highly-active antiretroviral therapy. *Lancet* **1998**;352:783–4.
26. Lucas GM, Eustace JA, Sozio S, Mentari EK, Appiah KA, Moore RD. Highly active antiretroviral therapy and the incidence of HIV-1-associated nephropathy: a 12-year cohort study. *AIDS* **2004**;18:541–6.
27. Wei A, Burns GC, Williams BA, Mohammed NB, Visintainer P, Sivak SL. Long-term renal survival in HIV-associated nephropathy with angiotensin-converting enzyme inhibition. *Kidney Int* **2003**;64:1462–71.
28. Szczech LA, Edwards LJ, Sanders LL, et al. Protease inhibitors are associated with a slowed progression of HIV-related renal diseases. *Clin Nephrol* **2002**;57:336–41.
29. Alpers CE. Light at the end of the TUNEL: HIV-associated thrombotic microangiopathy. *Kidney Int* **2003**;63:385–96.
30. del Arco A, Martinez MA, Pena JM, et al. Thrombotic thrombocytopenic purpura associated with human immunodeficiency virus infection: demonstration of p24 antigen in endothelial cells. *Clin Infect Dis* **1993**;17:360–3.
31. Hymes KB, Karpatikin S. Human immunodeficiency virus infection and thrombotic microangiopathy. *Semin Hematol* **1997**;34:117–25.
32. Sahud MA, Clater S, Liu L, Ero M, Harris K, Furlan M. von Willenrad factor-cleaving protease inhibitor in a patient with human immunodeficiency syndrome-associated thrombotic thrombocytopenic purpura. *Br J Haematol* **2002**;116:909–11.
33. Weiner NJ, Goodman JW, Kimmel PL. The HIV-associated renal diseases: current insight into pathogenesis and treatment. *Kidney Int* **2003**;63:1618–31.
34. Nochy D, Glotz D, Dosquet P, et al. Renal disease associated with HIV infection: a multicentric study of 60 patients from Paris hospitals. *Nephrol Dial Transplant* **1993**;8:11–9.
35. Beaufils H, Jouanneau C, Katlama C, Sazdovitch V, Hauw JJ. HIV-associated IgA nephropathy: a post-mortem study. *Nephrol Dial Transplant* **1995**;10:35–8.
36. Haas M, Kaul S, Eustace JA. HIV-associated immune complex glomerulonephritis with “lupus-like” features: a clinicopathological study of 14 cases. *Kidney Int* **2005**;67:1381–90.
37. Casanova S, Mazzucco G, Barbiano di Belgiojoso G, et al. Pattern of glomerular involvement in human immunodeficiency virus-infected patients: an Italian study. *Am J Kidney Dis* **1995**;26:446–53.
38. Tabechian D, Pattanaik D, Suresh U, Cohn SE, Nadasdy T. Lupus-like nephritis in an HIV-positive patient: report of a case and review of the literature. *Clin Nephrol* **2003**;60:187–94.
39. Gorriz JL, Rovira E, Sancho A, Ferrer R, Paricio A, Pallardo LM. IgA nephropathy associated with human immunodeficiency virus infection: antiproteinuric effect of captopril. *Nephrol Dial Transplant* **1997**;12:2796–7.
40. Walli R, Goebel FD, Demant T. Impaired glucose tolerance and protease inhibitors. *Ann Intern Med* **1998**;129:837–8.
41. Walli R, Herfort O, Michl GM, et al. Treatment with protease inhibitors associated with peripheral insulin resistance and impaired oral glucose tolerance in HIV-1-infected patients. *AIDS* **1998**;12:F167–73.
42. Friis-Moller N, Weber R, Reiss P, et al. Cardiovascular disease risk

- factors in HIV patients—association with antiretroviral therapy: results from the DAD study. *AIDS* **2003**; 17:1179–93.
43. Brown TT, Cole SR, Li X, et al. Antiretroviral therapy and the prevalence and incidence of diabetes mellitus in the multicenter AIDS cohort study. *Arch Intern Med* **2005**; 165:1179–84.
  44. Seaberg EC, Munoz A, Lu M, et al. Association between highly active antiretroviral therapy and hypertension in a large cohort of men followed from 1984 to 2003. *AIDS* **2005**; 19:953–60.
  45. Szczech LA, Gupta SK, Habash R, et al. The clinical epidemiology and course of the spectrum of renal diseases associated with HIV infection. *Kidney Int* **2004**; 66:1145–52.
  46. Daugas E, Rougier JP, Hill G. HAART-related nephropathies in HIV-infected patients. *Kidney Int* **2005**; 67:393–403.
  47. Kopp JB, Miller KD, Mican JA, et al. Crystalluria and urinary tract abnormalities associated with indinavir. *Ann Intern Med* **1997**; 127:119–25.
  48. Dieleman JP, Sturkenboom MC, Jambroes M, et al. Risk factors for urological symptoms in a cohort of users of the HIV protease inhibitor indinavir sulfate: the ATHENA cohort. *Arch Intern Med* **2002**; 162:1493–501.
  49. de Araujo M, Seguro AC. Trimethoprim-sulfamethoxazole (TMP/SMX) potentiates indinavir nephrotoxicity. *Antivir Ther* **2002**; 7:181–4.
  50. Herman JS, Ives NJ, Nelson M, Gazzard BG, Easterbrook PJ. Incidence and risk factors for the development of indinavir-associated renal complications. *J Antimicrob Chemother* **2001**; 48:355–60.
  51. Malavaud B, Dinh B, Bonnet E, Izopet J, Payen JL, Marchou B. Increased incidence of indinavir nephrolithiasis in patients with hepatitis B or C virus infection. *Antivir Ther* **2000**; 5:3–5.
  52. Casado JL, Moreno A, Sabido R, et al. A clinical study of the combination of 100 mg ritonavir plus 800 mg indinavir as salvage therapy: influence of increased plasma drug levels in the rate of response. *HIV Clin Trials* **2000**; 1:13–9.
  53. Daudon M, Estepa L, Viard JP, Joly D, Jungers P. Urinary stones in HIV-1-positive patients treated with indinavir. *Lancet* **1997**; 349:1294–5.
  54. Guery B, Katlama C, Bochet M, Deray G. Renal tolerance of combined treatment with foscarnet and indinavir. *Nephrol Dial Transplant* **1998**; 13:815–6.
  55. Sarcletti M, Petter A, Romani N, et al. Pyuria in patients treated with indinavir is associated with renal dysfunction. *Clin Nephrol* **2000**; 54:261–7.
  56. Dieleman JP, van Rossum AM, Stricker BC, et al. Persistent leukocyturia and loss of renal function in a prospectively monitored cohort of HIV-infected patients treated with indinavir. *J Acquir Immune Defic Syndr* **2003**; 32:135–42.
  57. van Rossum AM, Dieleman JP, Fraaij PL, et al. Persistent sterile leukocyturia is associated with impaired renal function in human immunodeficiency virus type 1-infected children treated with indinavir. *Pediatrics* **2002**; 110:e19.
  58. Boubaker K, Sudre P, Bally F, et al. Changes in renal function associated with indinavir. *AIDS* **1998**; 12:F249–54.
  59. Jaradat M, Phillips C, Yum MN, Cushing H, Moe S. Acute tubulointerstitial nephritis attributable to indinavir therapy. *Am J Kidney Dis* **2000**; 35:E16.
  60. Martinez F, Mommeja-Marin H, Estepa-Maurice L, et al. Indinavir crystal deposits associated with tubulointerstitial nephropathy. *Nephrol Dial Transplant* **1998**; 13:750–3.
  61. Tashima KT, Horowitz JD, Rosen S. Indinavir nephropathy. *N Engl J Med* **1997**; 336:138–40.
  62. Hanabusa H, Tagami H, Hataya H. Renal atrophy associated with long-term treatment with indinavir. *N Engl J Med* **1999**; 340:392–3.
  63. Chugh S, Bird R, Alexander EA. Ritonavir and renal failure. *N Engl J Med* **1997**; 336:138.
  64. Bochet MV, Jacquiaud C, Valantin MA, Katlama C, Deray G. Renal insufficiency induced by ritonavir in HIV-infected patients. *Am J Med* **1998**; 105:457.
  65. Duong M, Sgro C, Grappin M, Biron F, Boibieux A. Renal failure after treatment with ritonavir. *Lancet* **1996**; 348:693.
  66. Green ST, McKendrick MW, Schmid ML, Mohsen AH, Prakasam SF. Renal calculi developing de novo in a patient taking saquinavir. *Int J STD AIDS* **1998**; 9:555.
  67. Engeler DS, John H, Rentsch KM, Ruef C, Oertle D, Suter S. Nelfinavir urinary stones. *J Urol* **2002**; 167:1384–5.
  68. Brewster UC, Perazella MA. Acute interstitial nephritis associated with atazanavir, a new protease inhibitor. *Am J Kidney Dis* **2004**; 44:e81–4.
  69. Verhelst D, Monge M, Meynard JL, et al. Fanconi syndrome and renal failure induced by tenofovir: a first case report. *Am J Kidney Dis* **2002**; 40:1331–3.
  70. Coca S, Perazella MA. Rapid communication: acute renal failure associated with tenofovir: evidence of drug-induced nephrotoxicity. *Am J Med Sci* **2002**; 324:342–4.
  71. Creput C, Gonzalez-Canali G, Hill G, Piketty C, Kazatchkine M, Nochy D. Renal lesions in HIV-1-positive patient treated with tenofovir. *AIDS* **2003**; 17:935–7.
  72. Karras A, Lafaurie M, Furco A, et al. Tenofovir-related nephrotoxicity in human immunodeficiency virus-infected patients: three cases of renal failure, Fanconi syndrome, and nephrogenic diabetes insipidus. *Clin Infect Dis* **2003**; 36:1070–3.
  73. Schaaf B, Aries SP, Kramme E, Steinhoff J, Dalhoff K. Acute renal failure associated with tenofovir treatment in a patient with acquired immunodeficiency syndrome. *Clin Infect Dis* **2003**; 37:e41–3.
  74. Lee JC, Marosok RD. Acute tubular necrosis in a patient receiving tenofovir. *AIDS* **2003**; 17:2543–4.
  75. Saumoy M, Vidal F, Peraire J, et al. Proximal tubular kidney damage and tenofovir: a role for mitochondrial toxicity? *AIDS* **2004**; 18:1741–2.
  76. Rifkin BS, Perazella MA. Tenofovir-associated nephrotoxicity: Fanconi syndrome and renal failure. *Am J Med* **2004**; 117:282–4.
  77. Barrios A, Garcia-Benayas T, Gonzalez-Lahoz J, Soriano V. Tenofovir-related nephrotoxicity in HIV-infected patients. *AIDS* **2004**; 18:960–3.
  78. Peyriere H, Reynes J, Rouanet I, et al. Renal tubular dysfunction associated with tenofovir therapy: report of 7 cases. *J Acquir Immune Defic Syndr* **2004**; 35:269–73.
  79. Izzedine H, Hulot JS, Vittecoq D, et al. Long-term renal safety of tenofovir disoproxil fumarate in antiretroviral-naïve HIV-1-infected patients: data from a double-blind randomized active-controlled multicentre study. *Nephrol Dial Transplant* **2005**; 20:743–6.
  80. Zimmermann AE, Pizzoferrato T, Bedford J, et al. Tenofovir-associated acute and chronic kidney disease: a case of multiple drug interactions. *Clin Infect Dis* **2006**; 42:283–90.
  81. Izzedine H, Launay-Vacher V, Isnard-Bagnis C, Deray G. Drug-induced Fanconi's syndrome. *Am J Kidney Dis* **2003**; 41:292–309.
  82. Antoniou T, Raboud J, Chirhin S, et al. Incidence of and risk factors for tenofovir-induced nephrotoxicity: a retrospective cohort study. *HIV Med* **2005**; 6:284–90.
  83. Julg BD, Bogner JR, Crispin A, Goebel FD. Progression of renal impairment under therapy with tenofovir. *AIDS* **2005**; 19:1332–3.
  84. Gallant JE, Parish MA, Keruly JC, Moore RD. Changes in renal function associated with tenofovir disoproxil fumarate treatment, compared with nucleoside reverse-transcriptase inhibitor treatment. *Clin Infect Dis* **2005**; 40:1194–8.
  85. Mauss S, Berger F, Schmutz G. Antiretroviral therapy with tenofovir is associated with mild renal dysfunction. *AIDS* **2005**; 19:93–5.
  86. Izzedine H, Launay-Vacher V, Deray G. Renal tubular transporters and antiviral drugs: an update. *AIDS* **2005**; 19:455–62.
  87. Cihlar T, Ho ES, Lin DC, Mulato AS. Human renal organic anion transporter 1 (hOAT1) and its role in the nephrotoxicity of antiviral nucleotide analogs. *Nucleosides Nucleotides Nucleic Acids* **2001**; 20:641–8.
  88. Schuetz JD, Connelly MC, Sun D, et al. MRP4: a previously unidentified factor in resistance to nucleoside-based antiviral drugs. *Nat Med* **1999**; 5:1048–51.
  89. Izzedine H, Launay-Vacher V, Deray G. Antiviral drug-induced nephrotoxicity. *Am J Kidney Dis* **2005**; 45:804–17.
  90. Gutmann H, Fricker G, Drewe J, Toeroek M, Miller DS. Interactions

- of HIV protease inhibitors with ATP-dependent drug export proteins. *Mol Pharmacol* **1999**; 56:383–9.
91. Reid G, Wielinga P, Zelcer N, et al. Characterization of the transport of nucleoside analog drugs by the human multidrug resistance proteins MRP4 and MRP5. *Mol Pharmacol* **2003**; 63:1094–103.
  92. Gallant JE, Staszewski S, Pozniak AL, et al. Efficacy and safety of tenofovir DF vs stavudine in combination therapy in antiretroviral-naïve patients: a 3-year randomized trial. *JAMA* **2004**; 292:191–201.
  93. Squires K, Pozniak AL, Pierone G Jr, et al. Tenofovir disoproxil fumarate in nucleoside-resistant HIV-1 infection: a randomized trial. *Ann Intern Med* **2003**; 139:313–20.
  94. Schooley RT, Ruane P, Myers RA, et al. Tenofovir DF in antiretroviral-experienced patients: results from a 48-week, randomized, double-blind study. *AIDS* **2002**; 16:1257–63.
  95. Jones R, Stebbing J, Nelson M, et al. Renal dysfunction with tenofovir disoproxil fumarate-containing highly active antiretroviral therapy regimens is not observed more frequently: a cohort and case-control study. *J Acquir Immune Defic Syndr* **2004**; 37:1489–95.
  96. Izzedine H, Launay-Vacher V, Deray G. Fanconi syndrome associated with didanosine therapy. *AIDS* **2005**; 19:844–5.
  97. Crowther MA, Callaghan W, Hodsman AB, Mackie ID. Dideoxyinosine-associated nephrotoxicity. *AIDS* **1993**; 7:131–2.
  98. Morris AA, Baudouin SV, Snow MH. Renal tubular acidosis and hypophosphataemia after treatment with nucleoside reverse transcriptase inhibitors. *AIDS* **2001**; 15:140–1.
  99. Krishnan M, Nair R, Haas M, Atta MG. Acute renal failure in an HIV-positive 50-year-old man. *Am J Kidney Dis* **2000**; 36:1075–8.
  100. Angel-Moreno-Maroto A, Suarez-Castellano L, Hernandez-Cabrera M, Perez-Arellano JL. Severe efavirenz-induced hypersensitivity syndrome (not-DRESS) with acute renal failure. *J Infect* **2006**; 52:e39–40.
  101. Lalezari JP, Henry K, O'Hearn M, et al. Enfuvirtide, an HIV-1 fusion inhibitor, for drug-resistant HIV infection in North and South America. *N Engl J Med* **2003**; 348:2175–85.