# Role of Infection as a Risk Factor for Atherosclerosis, Myocardial Infarction, and Stroke

K. J. Mattila, V. V. Valtonen, M. S. Nieminen, and S. Asikainen

From Bayer Finland, Espoo; Department of Medicine, Divisions of Cardiology and Infectious Diseases, Helsinki University Central Hospital, Helsinki; and Department of Periodontology, Institute of Dentistry, University of Helsinki, Helsinki, Finland

An increasing body of evidence has linked infections to atherosclerosis and thrombosis. Herpesviruses cause atherosclerosis in experimental animals. Herpesviruses can also be detected in atherosclerotic lesions in humans. Cytomegalovirus may play a role in arteriosclerosis in transplanted hearts, and this virus, together with tumor suppressor protein p53, can be found in restenosis lesions following angioplasty. *Chlamydia pneumoniae* and dental infections are associated with coronary heart disease in cross-sectional and longitudinal studies, and preceding respiratory infections are associated with ischemic stroke. Infections may favor formation of atherosclerosis and thrombosis by elevation of blood levels of fibrinogen, leukocytes, clotting factor, and cytokines and by alteration of the metabolism and functions of endothelial cells and monocyte macrophages. Low-grade infections may also be one of the causes of the inflammatory reaction observed in atherosclerotic lesions and acute ischemic symptoms, reflected in elevated levels of C-reactive protein. These observations warrant further studies in this field.

The development of atherosclerosis in arterial walls, with complications such as chronic coronary heart disease (CHD), acute myocardial infarction (AMI), and ischemic stroke, has remained one of the leading causes of death and morbidity in Western industrialized countries. The classic risk factors for these disease entities do not explain all their clinical and epidemiological features. An increasing body of evidence suggests that infections also play a role, an idea proposed by Sir William Osler in the beginning of this century [1]. Inflammation, one of the causes of which may be low-grade infections, has been shown to be associated with atherosclerosis and acute coronary syndromes [2].

It should be emphasized that this "infection hypothesis" regarding cardiovascular diseases does not argue against the role of the classic coronary risk factors. Infections may in fact act through or in concert with the same elements as the classic risk factors, e.g., serum lipoproteins or thrombus formation.

The atherosclerotic process may begin in childhood, and as discussed later, it is possible that childhood infections are involved in this process; thus, infections and atherosclerosis may be tightly linked throughout human life.

This article reviews the recent evidence linking infections to various forms of atherosclerosis and its complications, with emphasis on the most recent findings in humans. Pure inflammatory reactions (quite arbitrarily defined) are not described

Received 31 March 1997; revised 16 October 1997.

Clinical Infectious Diseases 1998;26:719–34 © 1998 by The University of Chicago. All rights reserved. 1058–4838/98/2603–0027\$03.00 in detail, and some immunologic phenomena observed in patients with atherosclerosis-related diseases, like the presence of anticardiolipin antibodies, are considered beyond the scope of this review.

#### Atherosclerosis

Atherosclerosis is the common background behind acute myocardial infarction, unstable angina, and ischemic stroke. Therefore, it is important to elucidate the relation of infections to atherosclerosis; indeed, many different viral and bacterial infections have been linked with the disease.

# Viruses

Herpesviruses. The observation in the late 1970s that a herpes-type DNA virus of chickens (Marek's disease virus) causes atherosclerosis closely resembling that seen in humans meant a new emergence of the infection hypothesis regarding CHD. The atherosclerotic lesions developed also in normocholesterolemic virus-infected chickens; a cholesterol-rich diet changed the lesions from proliferative to fatty and fattyproliferative. The virus could be found in the arterial walls by immunofluorescence techniques [3]. Cultured chicken aortic smooth-muscle cells infected with this virus were also shown to contain higher amounts of cholesterol and other lipids than noninfected cells [4]. A twofold increase in cholesteryl ester synthetic activity and a significant reduction in lysosomal and cytoplasmic cholesteryl ester hydrolytic activity were observed in postmortem analysis of aortic wall specimens [4, 5]. What's more, Marek's disease virus-induced atherosclerosis could be

Reprints or correspondence: Dr. Kimmo Mattila, Bayer Finland, Suomalaistentie 7, 02270 Espoo, Finland.

prevented by administering herpesvirus from turkeys as a vaccine before the infection [6].

Span et al. [7] examined the effect of cytomegalovirus (CMV) on the development of atherosclerosis in rats. Increased adhesion of leukocytes to aortic intima was observed in CMV-infected normocholesterolemic rats, which also developed atherosclerotic lesions resembling those found in noninfected hypercholesterolemic rats. In hypercholesterolemic rats, CMV infection further increased the proportion of endothelial cells containing lipids, as compared with the proportion in hyper-cholesterolemic noninfected rats. These observations strongly suggest that herpesviruses can promote atherosclerosis in experimental animals.

These studies have been extended to humans. Herpesviruses are present in atherosclerotic arteries, as shown by Gyorkey et al. [8] in a study of patients with atherosclerosis undergoing cardiovascular surgery; by Benditt et al. [9] in a study of patients undergoing coronary bypass surgery; by Yamashiroya et al. [10] in a study of the coronary arteries of young trauma victims; and by Melnick et al. [11], who found the viruses in cultures of aortic cells and in the carotid arteries of 132 patients undergoing blood vessel surgery. Raza-Ahmad et al. [12] found histologic evidence of inflammation in 61% of coronary artery biopsy specimens taken during bypass grafting, and 45% of the samples were positive for herpes simplex virus type 2 antigens and 1% for herpes simplex virus type 1 antigens. What's more, there was a significant association between recent occurrence of cold sores, positivity for herpes simplex virus type 2, and inflammatory changes in the biopsy specimens.

The amount of herpesviruses has been correlated with the severity of the atherosclerotic process in the arterial tree. CMV nucleic acids could be demonstrated by PCR in 90% of samples taken from the abdominal aorta or femoral artery of CMV-seropositive patients undergoing vascular surgery, as compared with 53% of samples taken at autopsy (performed within 12 hours after death) from CMV-seropositive individuals with atherosclerosis of maximal grade I [13]. However, there seems to be no difference in the occurrence of CMV in atherosclerotic and uninvolved aortic segments within the arterial trees of the subjects studied; CMV was detected in up to 90% of the samples [14, 15].

A significantly higher prevalence of CMV antibodies and higher titers were observed among patients undergoing vascular surgery for atherosclerosis than among hyperlipidemic controls matched for age, race, and socioeconomic background [16]. It should be noted that this study is one of few comparing patients with atherosclerotic vessel disease with controls matched also for presence of hyperlipidemia. Sorlie et al. [17] analyzed antibodies to CMV and herpes simplex type 1 and 2 viruses in 340 matched case-control pairs. Cases had thickened carotid artery walls, as noted in ultrasonographic examinations. However, no statistically significant association was observed between carotid atherosclerosis and any of the viruses in multivariate analyses. The contribution of infections to atherosclerosis and restenosis was recently reviewed by Epstein et al. [18]. The above studies clearly indicate that CMV and other herpesviruses can be found in the arterial walls, which present a site of latency for these viruses. As stated by Hendrix et al. [13], one interpretation of these findings is that herpesviruses may be involved in the pathogenesis of atherosclerosis but that end-stage atherosclerosis may no longer be influenced by them. However, because of inflammation in the blood vessel wall, they could expose the individual to thrombotic complications.

Restenosis of the dilated vessel may occur in up to 50% of patients after coronary angioplasty. Recently, Speir et al. [19] found that 38% of restenosis lesions examined contained tumor suppressor protein p53 and that this correlated with the presence of CMV. Smooth-muscle cells from the lesion areas expressed CMV protein IE84 and contained high amounts of p53. Furthermore, CMV infection of cultured human smooth-muscle cells enhanced p53 accumulation, which in turn had a temporal association with IE84 expression. These results were given strong support by the observations of Zhou et al. [20]. In their prospective study, restenosis after coronary atherectomy occurred in 43% of CMV-antibody positive patients, as compared with only 8% of CMV-negative patients, a highly significant difference. It seems likely that CMV, via IE84-mediated inhibition of p53 function, is involved in the development of coronary restenosis.

The major factor limiting long-term survival after cardiac transplantation is the arteriosclerotic process taking place in the arteries of the transplanted heart, a phenomenon considered to represent chronic rejection. Practically all patients surviving for >1 year have vascular changes in the cardiac allograft at autopsy [21]. Morphologically, these changes consist of perivascular inflammation, focal myocytic necrosis, and concentric generalized intimal thickening [22].

Nucleic acids of CMV (together with those of herpes simplex virus and Epstein-Barr virus) can be found with in situ hybridization in coronary arteries of transplanted hearts [23, 24]. CMV infection has been associated with chronic rejection and arteriosclerosis in cardiac allografts in several [25–27] but not all clinical studies [28–30].

The carefully conducted study by Koskinen et al. [27] strongly suggests that CMV has a role in cardiac allograft arteriosclerosis. These investigators carried out a detailed analysis of consecutive endomyocardial biopsy specimens and coronary angiograms, taken regularly over 4 postoperative years from 53 heart transplant recipients. For the first 2 postoperative years, CMV infection was significantly associated with endothelial cell accumulation and thickened intima; the endothelial cell response subsided after this, but the intimal thickness continued to increase. In coronary angiography, CMV infection–associated acceleration of coronary narrowing was marked for 2–3 postoperative years, after which it subsided.

Still another form of cardiac allograft vasculopathy, distinct from the slowly progressing arteriosclerosis described above in that it leads to nonfunctioning of grafts within 1 year, has been reported by Paavonen et al. [31]. These investigators observed a strong inflammatory reaction with lymphocyte accumulation taking place in the subendothelial space in the intima, a phenomenon they referred to as endothelialitis. This reaction was also associated with CMV infection, and the investigators concluded that CMV may play a role in this form of arterial injury as well.

The immunosuppressed state of organ transplant recipients may of course make way for herpesviruses, and this should be taken into consideration when one interprets these results.

*HIV.* Paton et al. [32] described a series of eight HIVpositive males (five of whom were classified as having AIDS) whose autopsies revealed marked excentric atherosclerotic changes (up to 80%-90% obstruction) in the coronary arteries. These changes were disproportionate for the patients' young age (23–32 years) and lack of family history of coronary artery disease and other coronary risk factors (except for smoking by some of the individuals). The excentric type of lesions was different from the concentric type observed after cardiac transplantation. The authors concluded that viral infection, either by HIV or coexisting herpesviruses, had probably contributed to the development of the coronary lesions.

## Bacteria

Bacterial infections and intimal thickenings in the coronary arteries of infants. Intimal thickenings, composed mainly of smooth-muscle cell proliferations, can be observed in the coronary arteries even in infancy. These intimal thickenings are related to the well-known risk factors of CHD, such as male sex, place of birth, and family history of CHD [33]. It was noted as early as the 1940s that particularly thick intima layers in the coronary arteries could be found in the context of pneumonia [34].

This finding has been confirmed and examined in more detail by Pesonen et al. [35] and Kaprio et al. [33]. Their studies showed that preceding infections, mostly of bacterial origin, are associated with intimal thickening and an increased degree of luminal narrowing, even after adjustment for potential confounders such as age, gender, and family history of CHD. As the intimal thickenings are suspected to be precursors of future atherosclerotic lesions, these observations link infections with CHD from a very early age.

Kawasaki disease is presumed to have an infectious etiology. Occurring mostly in children aged  $\leq 2$  years, it carries sequelae into adulthood. The details of this disease are beyond the scope of this review, but it is common knowledge that some of the patients develop CHD at a young age [36, 37].

Chlamydia pneumoniae. Seropositivity for Chlamydia pneumoniae was significantly more common among patients

with AMI and chronic CHD than among random controls matched for age group, sex, and locality [38]. The association could not be explained by the classic coronary risk factors. Since this original observation, the association between *C. pneumoniae* and cardiovascular diseases has been confirmed and extended in several studies.

Individuals with CHD verified clinically with angiography or ultrasonography carry serological markers of chronic *C. pneumoniae* infection significantly more often than do matched random controls. These markers include more prevalent and higher titers of *C. pneumoniae* antibodies [39–43] (positive serology is associated with a 2 to 7 fold risk of CHD) and more prevalent circulating immune complexes containing *C. pneumoniae* [44]. The latter study indicated that many patients with atherosclerosis seem to have chlamydial components in their blood circulation.

In the study of Thom et al. [40], the association between CHD and *C. pneumoniae* seemed to be limited to smokers. Furthermore, when individuals with normal coronary angiograms were used as the control group, the association was no longer statistically significant. However, the association between positive *C. pneumoniae* serology and CHD remained significant even after adjustment for the effect of smoking and other potential confounders in all other studies cited above.

Dahlén et al. [43] found that the combination of male sex, HLA (human leukocyte antigen) DR II genotype 13a or 17, elevated Lp(a) levels (>120 mg/L), and IgG titers of >256 (positive *C. pneumoniae* serology) was very strongly associated with CHD (this combination was observed in 14 of 29 patients with CHD and in 1 of 27 controls). These findings suggest that genetic susceptibility and an infectious factor are involved in *C. pneumoniae*–associated atherosclerosis, a scenario resembling that occurring in the pathogenesis of reactive arthritis.

In 1992, C. pneumoniae was detected in coronary arterial fatty streaks and atheromatous plaques with use of electron microscopy and/or immunoperoxidase staining [45]. Kuo et al. found C. pneumoniae in coronary artery atheromas by means of immunocytochemistry, PCR, and electron microscopy; some 40%-55% of the samples studied were positive by at least one of the techniques [46]. The investigators extended their studies by showing C. pneumoniae in fatty streaks and fibrous plaques, where the organism was found both in macrophages and in smooth-muscle cells [47]. In the material of Campbell et al. [48], C. pneumoniae was found in 20 of 38 specimens from atherosclerotic vessels, and there was a trend toward the occurrence of C. pneumoniae more frequently in restenosis (following coronary angioplasty) lesions than in native lesions. In another study C. pneumoniae was observed in 5 of 5 carotid artery endarterectomy specimens with atherosclerotic changes, and it was not found in any of the 13 normal specimens [49].

Furthermore, the same investigators have demonstrated that *C. pneumoniae* cannot be detected in nonatherosclerotic areas

of the arteries of young (15–34 years old) individuals [50]. This finding was confirmed by Muhlestein et al., who found evidence of *C. pneumoniae* in 79% of atherosclerotic coronary tissue specimens but in only 4% of nonatherosclerotic specimens [51]. On the other hand, Weiss et al. [52] found *C. pneumoniae* in only one of 58 atherosclerotic coronary specimens they investigated.

Blasi et al. found *C. pneumoniae* in aortic aneurysm samples from 26 of 51 subjects undergoing abdominal aortic surgery [53]. This observation has been confirmed by Juvonen et al. [54], who found chlamydial lipopolysaccharide (LPS) and antigens in abdominal aortic aneurysms but not in control samples from healthy aortic tissue.

Of note, in the (quite small) materials studied, the presence of *C. pneumoniae* in atherosclerotic vessels has not been associated with positive *C. pneumoniae* serology [48]. On the contrary, in the study of Puolakkainen et al. [55], the microbe was found more often in the vessels in patients with low titers of IgG to *C. pneumoniae*. In this study, reactivities with 42-kD and 52-kD proteins were associated with the microbe's presence in the vessel wall. Serology therefore poorly reflects the *C. pneumoniae*–associated arterial disease status. The above results should be taken into consideration when further studies in this field are planned.

In 1995 *C. pneumoniae* was finally isolated from a coronary atherosclerotic lesion [56]. It is interesting that preliminary findings suggest that a positive *C. pneumoniae* serology is also associated with hypertension [57].

Thus, several lines of evidence link *C. pneumoniae* to atherosclerosis. The studies cited above, however, cannot definitively answer whether the microbe really is a causative organism in the atherosclerotic process or whether it is just a bystander that locates in diseased segments of the arterial tree.

#### **Dental Infections**

Caries and periodontal disease are the most common dental diseases. Gram-positive bacteria, especially *Streptococcus mutans*, are involved with the etiology of caries [58], whereas gram-negative bacteria, including *Porphyromonas gingivalis*, *Actinobacillus actinomycetemcomitans*, and *Prevotella intermedia*, are involved with that of periodontitis [59]. The host's reaction to bacterial agents plays an important role in the destructive process in periodontitis [60]. The course of caries and periodontitis is usually chronic, but occasionally acute complications occur, such as endodontal infections and periodontal abscesses as well as various odontogenic infections in the mandible, maxilla, and soft tissues in the neck area.

The prevalence of periodontitis increases with the age of the subjects studied; prevalence rates have varied between studies. Burt concludes in his review that the prevalence is quite small [61]. However, in a detailed survey carried out in Finland in the late 1970s [62], periodontal tissues were healthy in only

4% of subjects, and the prevalences of moderately severe and severe periodontitis were 50% and 20%, respectively.

Mattila et al. [63] studied the relation of dental infections to coronary atherosclerosis in 100 patients with proven CHD by relating a score based on pantomographic findings (the sum of the number of periapical lesions, lesions caused by tertiary caries, vertical bone pockets, and furcation and pericoronitis lesions), assessed blindly by a single dentist, to the extent of coronary atheromatosis. This was scored on the basis of coronary angiographs, also assessed blindly by a single radiologist. The pantomography index was significantly higher among those with advanced coronary atherosclerosis, and this association remained significant in multivariate analysis taking into account the effect of age, serum lipids, smoking, hypertension, social class, and other confounders [63].

It could be argued that atherosclerosis is the *cause* and not the consequence of dental infections; individuals with coronary atherosclerosis could also have compromised arterial circulation in the periodontium. The recent study of Adams et al. [64], however, strongly argues against this explanation. The investigators performed ultrasonographic examination of the carotid arteries in 350 consecutive patients with angiographically verified coronary atherosclerosis. Intima-media thickness in the carotid arteries was correlated with the extent of coronary atherosclerosis, but this correlation, although statistically significant, was considered clinically unimportant (r = 0.29;  $r^2 < 0.1$ ). Thus, it is unlikely that there would be any strong correlation between the coronary and gingival arteries either. Rather, the finding suggests that patients with CHD do not necessarily have severe atherosclerotic changes in the arteries in the region of the head.

Thus, dental infections represent another type of chronic bacterial infection that is quite strongly associated with athero-sclerosis.

# Acute Myocardial Infarction and Unstable Angina Pectoris

These acute coronary events are characterized by thrombus formation in a diseased coronary artery segment. Both viral and bacterial infections have been observed in conjunction with these entities as well.

#### Viruses

It is likely that some of the cases diagnosed as acute myocardial infarctions may in reality be focal myocarditis [65-67]. Individuals with acute myocardial infarction sometimes have evidence of recent viral infections, especially with coxsackie B virus. Controlled studies in this field have yielded conflicting results [68-75]. The epidemic nature of viral diseases, the very large number of infective agents, and the great variation in antibody response between individuals make this topic difficult to investigate in clinical studies. The relatively small sample sizes indicate the possibility of a type II error as well.

As CMV and inflammation have been implicated in the pathogenesis of coronary artery disease, Kol et al. [76] investigated the role of CMV replication in atherosclerotic plaque instability. They examined coronary atherectomy specimens from 20 patients with unstable angina pectoris. However, none of the specimens tested showed a positive signal for markers of early replication, thus arguing against a role for CMV replication in unstable angina pectoris.

A preceding "influenza-like illness," defined by symptoms of upper respiratory tract infection (i.e., nasal congestion, rhinorrhea, sore throat, head cold, or cough, with or without fever), was significantly associated with acute myocardial infarction in three studies specifically designed to study the phenomenon [35, 75, 77]. Lack of association has been noted in other investigations originally not designed to study this particular phenomenon [67, 68]. This syndrome occurred 1–5 weeks before the infarction in  $\sim 20\%$ –30% of patients (a 2.5-fold prevalence in comparison with that among random controls or patients with chronic CHD in a "cold" phase). So far, no association has been found between this syndrome and a positive viral or bacterial serology or dental infections. It is noteworthy, however, that the above symptoms again refer to inflammation, known to occur in acute cardiovascular events.

On the other hand, influenza epidemics are associated with increased numbers of deaths due to CHD, especially in younger age groups [78]. This could be explained by the fact that levels of fibrinogen and factor VIII clotting activity are higher in winter, causing a "hypercoagulabile" state that in turn is strongly correlated with an elevated neutrophil count, elevated C-reactive protein (CRP) level, and self-reported cough [79].

In accordance with the above, Jousilahti et al. [80] recently published a report on a 13-year follow-up of >19,000 randomly selected individuals from eastern Finland (individuals with a history of CHD or stroke were removed from the cohort). The investigators defined chronic respiratory infection as occurrence of symptoms of chronic bronchitis. In their large amount of carefully analyzed data, symptoms of chronic respiratory infection predicted coronary death with a risk ratio of ~1.5. Adjusting the result for smoking, cholesterol level, and systolic blood pressure decreased the risk somewhat, but it remained statistically significant (as did the risk for developing CHD).

## Chlamydia pneumoniae

As mentioned before, *C. pneumoniae* seropositivity was significantly more common among patients with AMI and chronic CHD than among their matched random controls. Two-thirds of these patients with AMI showed seroconversion in EIA with LPS antigen, core-deficient Re-LPS, purified from *Salmonella minnesota* mutant (as compared with only one patient of the other groups combined). This Re-LPS shares with chlamydial LPS a major carbohydrate antigen (alpha-2-4 linked keto deoxyoctonate [KDO]-disaccharide). Chlamydial LPS has an additional epitope, alpha-2-8 linked KDO-disaccharide. A positive reaction was observed only in assays containing this latter epitope. No reaction was observed, for example, with LPS of *Yersinia enterocolitica* or with serum from patients known to have other types of circulating immune complexes [81, 82].

*C. pneumoniae*-specific immune complexes were detected significantly more often in patients with AMI (57%) than in random controls (12%) and patient controls with diseases characterized by circulating immune complexes (10%) [82]. Immediately after the AMI, most of the circulating *C. pneumoniae*-specific immune complexes were detectable with the IgM-capture EIA method. On the contrary, LPS capture detected most immune complexes 1 month later. This shift in the composition of these immune complexes was not observed in the samples taken from the random controls.

These results suggested that after AMI, the immune complexes were formed in chlamydial LPS excess, which then converted into IgM antibody excess. The reason for and significance of this interesting finding are not known.

Cook et al. [83] measured IgG, IgM, and IgA *C. pneumoniae* antibodies in 1,874 patients admitted to a hospital in Birmingham, England over a 2-year period. Serological markers of both acute and chronic *C. pneumoniae* infection were significantly more common among patients admitted with unstable angina, AMI, or ischemic stroke than in patients admitted with other conditions. It is noteworthy that the 2-year-long recruitment period in this study excludes the possibility that a rapid start or cessation of a *C. peumoniae*-associated epidemic would have influenced the differences between the groups. Furthermore, the results were obtained by comparing the patients with hospital controls, among whom factors that might increase the prevalence of chlamydial infections, like smoking and alcohol abuse, are probably more common than among random controls.

Positive C. peumoniae serology is associated with cardiovascular disease in follow-up studies as well. In the Helsinki Heart Study, after adjustment for the effect of the "classic" coronary risk factors (age, smoking, systolic blood pressure, the ratio of high-density lipoprotein [HDL] to total cholesterol), the presence of C. pneumoniae-specific immune complexes and/or an elevated IgA antibody titer 3–6 months before clinical signs of CHD was associated with a >2-fold risk of developing CHD [84]. As all individuals included in that study had to be free from CHD at the time of enrollment, the conclusions exclude the possibility that C. pneumoniae antibodies or immune complexes would represent only a false-positive result, due to infarcted myocardium, in the assays used. Preliminary results also suggest that a positive C. pneumoniae serology and an elevated level of CRP, when occurring in conjunction in patients with unstable angina, accurately predict future ischemic events [85].

In a 7-year follow-up of diabetic and nondiabetic subjects in two areas in Finland, Miettinen et al. [86] observed that *C. pneumoniae* seropositivity was associated with future CHD events in nondiabetic subjects in eastern Finland. However, no association could be observed in diabetics and in individuals in western Finland. The reasons for this kind of finding were unknown, but the authors speculated that diabetes is so strong a risk factor that it could have masked the effect of weaker risk factors.

As smoking exposes individuals to respiratory infections and cardiovascular diseases, smoking is potentially an important confounder in the link between *C. pneumoniae* and cardiovascular disease. In most studies, smoking has not been associated with a positive *C. pneumoniae* serology or, at the least, the *C. pneumoniae*–CHD association has remained significant in multivariate analyses controlling for the effect of smoking. In a study of Thom et al. [40] comparing patients with angiographically verified CHD with matched controls, a *C. pneumoniae*–CHD association could be observed only among "ever smokers."

Two studies have specifically addressed the relation of positive *C. pneumoniae* serology and smoking. Hahn and Golubjatnikov [87] investigated 365 outpatients with respiratory illness. Current smokers were significantly more likely than nonsmokers to have a *C. pneumoniae* antibody titer of  $\geq 1:128$ , and this titer category was positively correlated with current smoking.

Karvonen et al. [88] analyzed the relation of smoking to *C. pneumoniae* seropositivity in a large number of individuals (2,346). Overall, "ever smokers" had a 50% greater risk of *C. pneumoniae* IgG seropositivity. Ex-smokers and current smokers had little difference in this respect, suggesting that "ever smokers" should be handled as a single group in future studies.

Thus, in large series of individuals, smoking seems to increase the risk of developing chronic C. pneumoniae infection. It should be noted, however, that in the study of Hahn and Golubjatnikov [87], the individuals studied were younger (mean age, 34 years) than in most clinical studies on C. pneumoniae and CHD. In addition, the individuals were recruited from among patients with an ongoing respiratory illness (whereas random or healthy controls were used in most other studies). Looking at the odds ratios for C. pneumoniae seropositivity according to age groups in the study of Karvonen et al. [88], one can see that the findings in their study were more evident among ex-smokers and younger individuals, especially those younger than 36 years. Thus, the association between smoking and C. pneumoniae may have been at least partly due to the fact that smoking exposes individuals to a chronic C. pneumoniae infection at a younger age.

More important, even if smoking does expose individuals to chronic *C. pneumoniae* infection, this does not exclude the possibility that the microbe is involved in the pathogenesis of coronary artery disease. As Hahn and Golubjatnikov [87] themselves pointed out, chronic *C. pneumoniae* infection could have this effect, irrespective of the possibility that smoking has exposed the individual to the infection.

In the above-mentioned study of Karvonen et al. [88], *C. pneumoniae* seropositivity was more common among males than females, and this could not be explained by the more common smoking among males. This is in keeping with the male-female ratio of the prevalence of CHD. On the other hand, the prevalence of *C. pneumoniae* seropositivity was higher in southwestern than eastern Finland in their second study, a finding not in keeping with the higher prevalence of CHD in eastern Finland [89].

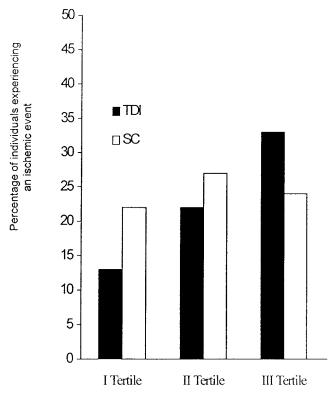
## **Dental Infections**

Dental infections were significantly associated with CHD in a case-control study of 100 patients with AMI and 102 random controls matched for locality, sex, and age range [90]. In this study, the severity of dental infections was graded by dentists using two scores. One score ("TDI") was based on nonblinded, systematic, clinicoradiological scoring of the severity of caries (0-3), periodontitis (0-3), periapical lesions (0-3), and pericoronitis (0-1) in the participants. The other comprised blind assessment of pantomographs, as described earlier.

The prevalence of edentulousness was higher in the AMI and chronic CHD groups than among the random controls. Multivariate analyses suggested that the association was independent of the effect of smoking, social class, and other potential confounders. The number of missing teeth, which may be regarded as an indirect indicator of dental infection experience, was related to the prevalence of CHD among 1,384 men aged 45–64 years living in different regions of Finland. In a multivariate analysis controlling for the effect of smoking, hypertension, educational level, and age, the number of missing teeth was significantly associated with CHD [91].

DeStefano et al. [92] have published the largest study in this field: a 14-year follow-up of 9,760 individuals who were free of CHD at enrollment and had undergone a dental examination at the beginning of the follow-up. Dental infections were quantitated by the number of decayed teeth, periodontal disease classification, and a periodontal index; an oral hygiene index was employed to indicate the degree of dental health. The results showed that among all subjects studied, after careful adjustment for several potential confounders, periodontitis was associated with a 25% increase in CHD risk. Among men younger than 50 years (most of the individuals in the abovementioned studies), this excess risk was 70%. However, the oral hygiene index was also associated with excess CHD risk and total mortality, and edentulousness was associated with excess total mortality.

The mechanism linking edentulousness with CHD in several studies remains speculative. Likewise, despite the large amount of data, long follow-up, and excellent statistics in the study by DeStefano et al., the truly independent role of dental infections in the development of CHD remains unclear.



**Figure 1.** Percentage of individuals experiencing an ischemic event during follow-up in the three tertiles of Total Dental Index (TDI) and Social Class (SC). Test for linear trend: TDI, P = .01; SC, P = not significant.

The patients with CHD in the two Finnish studies described [63, 75] have been followed for subsequent ischemic events. Among the 214 patients enrolled, 52 met the end-point criteria (fatal and nonfatal AMI, episodes of unstable angina) during the 7 years of follow-up. The severity of dental infections (i.e., the above-described "TDI") was significantly associated with the risk of ischemic events both in univariate analysis and in the Cox model, accounting for the effect of age, enrollment smoking status, social class, blood lipids, and other confounders [93].

As can be seen in figure 1 (based on the results of Mattila et al. [93], previously unpublished), the risk of an ischemic event increased from the lowest to the highest TDI tertile in a completely linear fashion, whereas no relation was observed between the event risk and social class.

Thus, the severity of dental infections seems to predict future CHD events in individuals with and without CHD at enrollment. It is clear that in these kinds of studies there is a risk of bias caused by lifestyle-related factors, such as diet and health care practices. A causal relationship is also possible, however, and further studies are indicated in this field.

## Helicobacter pylori

Patel et al. [42] compared men with electrocardiographic evidence of CHD with men whose electrocardiograms were normal, with regard to *Helicobacter pylori* and *C. pneumoniae* seropositivity. Serological evidence of these infections was associated with odds ratios of 3.8 and 3.1, respectively, for CHD after adjustment for various confounders. Martîn-de-Argila et al. [94] published their findings with regard to 101 CHD patients and 68 healthy controls of similar age and socioeconomic status. The *H. pylori* seropositivity rate was 84% among the patients with CHD vs. 58% among the controls, a highly significant difference. Likewise, in the material published by Morgando et al. [95], *H. pylori* was again associated with CHD, especially among those younger than 60 years of age.

However, Murray et al. [96] could not associate *H. pylori* seropositivity with CHD, high blood pressure, plasma viscosity, total cholesterol level, and social class for randomly selected subjects aged 25–64 years (1,182 men and 1,198 women). A weak negative association was found between *H. pylori* and fibrinogen and between *H. pylori* and HDL in women. A lack of association between *H. pylori* and CHD was reported also by Sandifer et al. [97] and Delaney et al. [98]. Furthermore, the only prospective study on *H. pylori* seropositivity and the risk of CHD in elderly individuals also failed to show any association between the infection and CHD [99].

Like *C. pneumoniae, H. pylori* was associated with hypertension and body mass index in another study, of 162 individuals [100].

#### **Enterobacterial Infections**

Enterobacterial common antigen is an antigenic component of all Enterobacteriaceae organisms, except defective mutants. It is not present in other gram-positive or gram-negative organisms [101]. Among the patients with AMI in the aforementioned study by Mattila et al. [75], 15 (38%) had at least fourfold increases in the level of enterobacterial common antigen antibodies; in comparison, one subject in the population control group and none in the chronic CHD group had a diagnostic rise in antibody level. Significant elevations in levels of IgG, IgA, and IgM anticardiolipin antibodies were also observed in the AMI group, and these two reactions were closely associated. The etiology and significance of these findings are unknown.

#### **Cerebral Infarction**

The infections observed in patients with cerebral infarction have mostly been of bacterial origin. It has long been known that infective endocarditis is complicated by cerebral infarction in up to 15% of cases [102]. Another well-known association has been that with *Mycoplasma pneumoniae* [103]. Observational studies have shown that some 5%-10% of patients with bacteremic infections but without endocarditis or CNS infection develop stroke within a few weeks after the onset of bacteremia [102, 104]. Uza reported that bacteriuria was almost six 726

times more common among patients with ischemic cerebrovascular disease than among age-matched controls [105].

In the case-control study of Syrjänen et al., febrile infection was associated with cerebral infarction with a relative risk of 9 (CI, 2.2-80), and the most common preceding infections were upper respiratory infections [106]. These associations remained significant in multivariate analyses controlling for the effect of other risk factors of cerebral infarction. The investigators additionally showed that young (<50 years old) male patients with ischemic stroke had significantly more severe dental infections than did the controls [107].

The association between recent infections and stroke was confirmed by Grau et al. [108], who compared 197 patients who had ischemic strokes with 197 random controls matched for age, sex, and area of residence. A preceding infection carried an increased risk for stroke, with an odds ratio of 4.5 (95% CI, 2.1–9.7); likewise, a febrile illness in the patient's history was associated with cerebral infarction, with an odds ratio of 7 (95% CI, 2.5–20). Again, the preceding infections in the stroke patients were upper respiratory infections of bacterial origin, and adjusting for the effect of potential confounders did not diminish the above associations.

Cook et al. [83] investigated the association between *C. pneumoniae* and various arterial thrombotic diseases, including ischemic cerebral syndromes. Serological markers of both acute and chronic *C. pneumoniae* infection were significantly more common in these patients than in hospital controls, and the magnitude of these differences was even bigger than that between patients with CHD and controls.

This finding was confirmed by Wimmer et al. [109], who showed that positive *C. pneumoniae* IgA antibody levels and *C. pneumoniae* immunocomplexes were associated with ischemic cerebral attacks after adjustment for age, sex, hypertension, and migraines, with an odds ratio of 2.2.

# Infection, Atherosclerosis, and Infarction: Possible Mechanisms of Action

The idea of infection as a risk factor for myocardial infarction may at first seem strange. However, there are several mechanisms that could mediate the effect of infections on both atherogenesis and thrombogenesis. For example, infections can evoke cytokine production; cytokines in turn probably play an important role in the pathogenesis of atherosclerosis [110].

## Viruses

At the least, herpes simplex virus type 1, adenovirus type 7, the measles and mumps viruses, poliovirus type 1, echovirus type 9, and parainfluenza type 3 viruses are capable of infecting human endothelial cells in culture [111]. Infection with herpes simplex virus type 1, poliovirus, and adenoviruses has been shown to increase granulocyte adherence to human endothelial

cells [112]. Adherent granulocytes can release toxic products (especially oxygen radicals) capable of damaging the endothelium after endothelial exposure to immune complexes, endotoxin, or activated complement [113-115]. Immune complexes in turn induce production of tissue factor, the initiator of the extrinsic coagulation pathway, in human endothelial cells [116]. Infection of human endothelial cells with herpes simplex virus type 1 leads to induction of immune complex receptors [117], enhanced thrombin generation and platelet binding [118], and decreased endothelial-cell plasminogen activator inhibitor [119]. Herpes simplex virus type 1 does not require productive infection to induce tissue factor in human umbilical vein endothelial cells [120]. Furthermore, herpesviruses can make the vascular endothelium more prone to thrombus formation by causing loss of surface heparans and thrombomodulin, a decrease in prostacyclin synthesis, and generation and release of tissue factor [121].

In human arterial smooth-muscle cells, this virus leads to accumulation of saturated cholesteryl esters and triglycerides (partly due to decreased cholesteryl ester hydrolysis) and a reduced capacity to produce prostacyclin [5]. Besides herpes simplex virus, CMV, at the least, is capable of replicating in human arterial smooth-muscle cells [122].

## Bacteria

Many studies of the interactions between bacterial products and host tissue have involved LPS, a structural component of gram-negative bacteria. Although less thoroughly studied, components of gram-positive bacteria have similar properties.

LPS influences endothelial cell function, even without induction of denuding endothelial damage. It enhances the synthesis of IL-1 in human monocytes and endothelial cells and of TNF in human endothelial cells. These substances induce procoagulant activity in human vascular endothelium and are known to increase the adherence of polymorphonuclear leukocytes and monocytes to it [123–126]. LPS stimulates endothelial production of platelet activating factor [127] and induces release of tissue factor [128, 129], plasminogen activator inhibitor [130, 131], and von Willebrand factor antigen [132] from human endothelial cells. Furthermore, LPS enhances production of the factor stimulating growth of granulocytes and macrophages and the colony stimulating activity in cultured human endothelial cells [133].

*C. pneumoniae* is able to multiply and cause a persistent infection in human endothelial cells [134]. It also induces expression of endothelial-leukocyte, intracellular, and vascular cell adhesion molecules in human endothelial cells [135].

LPS has several effects on monocyte-macrophages. LPS of *Escherichia coli* remains within macrophages [136]. LPS stimulates secretion of type-beta transforming growth factor in human monocytes [137] and stimulates secretion of growth factors for smooth muscle cells and fibroblasts and the expression

of c-sis proto-oncogene coding for one of the platelet-derived growth-factor chains in human monocytes [138–140], together with induction of IL-2 receptors on monocytes. IL-2 then augments generation of reactive oxidative intermediates and their cytotoxic activity in monocytes [141].

Monocyte adhesion to blood vessel endothelium and lipid metabolism of monocyte-macrophages are important factors in atherogenesis. As mentioned above, LPS enhances monocyte adherence to endothelium via stimulating the production of IL-1 and TNF in human monocytes and endothelial cells [142, 143] and, at least in animal preparates, by causing increased endothelial-cell turnover. The latter is also among the factors increasing monocyte adherence [144]. Monocytes are also involved in the indirect activation of endothelium by endotoxin, which takes place during the neutralization process of LPS. LPS activates monocytes via cell membrane CD14, leading to secretion of TNF and IL-1, which in turn activate endothelium [145].

Besides this, LPS induces secretion of factors chemotactic to monocytes in human arterial smooth muscle cells and macrophages [146]. Repeated exposure of monocytes to endotoxin augments the subsequent release of inflammatory mediators from monocytes [147]. This can be observed clinically, as shown by Schäfer et al. [148]. They showed that patients with liver diseases had elevated plasma endotoxin levels and that monocytes of these patients released increased amounts of bioactive TNF, as compared with such activity in healthy controls.

*C. pneumoniae* can grow inside human peripheral blood mononuclear cells, and it induces a cytokine response (with TNF, IL-1, IL-6, and IFN) in these cells [135]. Unstimulated and LPS-stimulated monocytes from patients with periodontitis, at least those who develop periodontitis at a relatively young age, release higher concentrations of IL-1 than do those from subjects with few or no dental infections [149].

An elevated granulocyte count is an independent risk factor for CHD, and leukocyte count correlates with the extent of angiographically assessed severity of coronary atherosclerosis [150]. As mentioned above, LPS significantly increases adherence of neutrophils to human endothelial cells. LPS activates granulocytes to increase the release of oxygen metabolites. Activated granulocytes in turn can damage endothelial cells via lysosomal enzymes, oxygen radicals, elastase, and other toxic substances in vitro [114, 115, 151–154].

Hydrogen peroxide stimulates the synthesis of platelet activating factor in endothelium and induces endothelial cell–dependent neutrophil adhesion [155]. LPS directly enhances production of platelet activating factor in neutrophils [156]. On the other hand, platelet activating factor enhances neutrophil responses, e.g., elastase release and aggregation, to various stimuli [157].

Besides its direct effects, LPS also activates granulocytes by stimulating monocytes to excrete granulocyte-activating mediators [158] and platelet-derived growth factor, also known to promote granulocyte activation [159].

In vivo investigations have shown that hydrogen peroxide production by neutrophils in patients with bacterial infection is increased [160], as is neutrophil elastase production in healthy volunteers given minute amounts of LPS [161]. Stimulated peripheral neutrophils of adult patients with periodontitis have been reported to have a >2-fold higher release of free oxygen radicals in comparison with those of individuals without periodontitis [162, 163]. Individuals with periodontal disease also have significantly elevated levels of fibrinogen and leukocytes, and these differences were not explained by factors like smoking or social class in the study of Kweider et al. [164].

The fate of LPS that invades the human body is linked with lipoproteins. All major lipoprotein classes in the blood bind LPS in direct proportion to their cholesterol concentration, and low-density lipoprotein (LDL) cholesterol binds most LPS in human plasma [165]. Lipoprotein-binding protein is also involved in the processing of LPS, by catalyzing the movement of LPS from endotoxin micelles to soluble CD14 (sCD14) and to plasma lipoproteins [166]. The LPS-sCD14 complexes can directly activate endothelium.

Injection of LPS into experimental animals leads to a hyperlipidemic response, which is considered a host-defense mechanism against this compound [167, 168]. The magnitude of the increase in cytokine production produced by endotoxin is greater if hypolipidemia is induced in the experimental animals [169].

In the studies of Morel et al. [169], neither LPS nor LDL cholesterol alone showed cytotoxicity to relatively quiescent human endothelial cells. However, LPS induced endothelial cells to produce free radicals, which oxidize LDL cholesterol. Oxidized LPS–LDL cholesterol complex then could enter the endothelial cells via the scavenger receptor, and this led to clearly diminished survival of endothelial cells. The toxicity could be inhibited by blockers of acetyl-LDL cholesterol scavenger receptor.

Brand et al. [170] showed that oxidized LDL cholesterol alone did not induce tissue factor expression in monocytes but it significantly enhanced tissue factor expression induced by LPS. LPS–LDL cholesterol complex can enter macrophages via the apolipoprotein B/E receptor [165]. After this, the complex affects the expression of scavenger receptor activity during monocyte differentiation in vitro [165] and activates monocytes to oxidize LDL cholesterol, making it toxic to other cells [171].

LPS causes increased cholesteryl ester synthesis and accumulation in human macrophages [172]. Analogous with the fact that HDL cholesterol (unlike LDL cholesterol) complexed with LPS is not toxic to human endothelial cells, HDL cholesterol inhibits the LPS-mediated increase in oxidative metabolism and lysozyme release of granulocytes [173].

Clinical studies have shown that both viral and bacterial infections cause significant changes in serum lipoprotein levels. In the study of Sammalkorpi et al. [174], a reduction in HDL related to the severity of the infection. The above-mentioned observations link bacterial compounds and infections to the levels of HDL and LDL cholesterol and the HDL/LDL cholesterol ratio, powerful predictors of CHD.

cholesterol to LDL cholesterol was observed. The changes were

Microbial compounds may also favor thrombus formation. *Staphylococcus aureus* has been shown to be as strong a stimulus for release reaction of human platelets as collagen, thrombin, and adrenalin [175]. Furthermore, *Fusobacterium necrophorum* and *Streptococcus pyogenes* have been shown to aggregate human platelets in vitro [176, 177]. *Streptococcus sanguis*, ubiquitous in dental plaque, can aggregate human platelets [178]. Miragliotta et al. [179] showed that *H. pylori* has tissue-factor-like procoagulant activity that could activate the extrinsic pathway of blood coagulation.

LPS can induce monocytes to aggregate platelets [180] and to produce factor VII activity in the presence of lymphocytes [181]. LPS induces generation of fast-acting inhibitor of plasminogen activator from human monocytes and endothelial cells. This has been demonstrated in experimental animals, in human preparates in vitro, and in patients with septicemia. The phenomenon can be induced both by LPS itself and via the action of IL-1 [130, 131, 182].

Furthermore, LPS can activate Hageman factor, suggesting that LPS can initiate the intrinsic pathway of coagulation [183]. The profound effects of LPS were clearly shown by Suffredini et al. [161], who gave minute amounts (4 ng/kg) of LPS to normal subjects. LPS induced a fourfold rise in the level of von Willebrand factor antigen and a clear "procoagulant state," characterized by an increase in plasminogen-activator inhibitor activity and diminished t-PA (tissue-type plasminogen activator) activity.

A procoagulant state, which may persist weeks after both viral and bacterial infections, has also been observed in clinical studies. This has been shown by measuring  $\beta$ -thromboglobulin, spontaneous aggregation of platelets, and platelet responsiveness to various stimuli [184, 185]. Likewise, elevations in fibrinogen levels after both viral and bacterial infections were observed in the 1970s [186]. Patients with severe dental infections seem to have elevated levels of von Willebrand factor antigen [187]; IgG, IgM, and IgD; and circulating immune complexes [188]. Ameriso et al. [189] explored the potential mechanisms mediating the association between infections and cerebral infarction. They compared cerebral infarction patients with (n = 17) and without (n = 33) a preceding infection, with regard to immunohematologic characteristics. Patients with a preceding infection had significantly higher levels of fibrin Ddimer and fibrinogen (studied within 2 days) and enhanced cardiolipin immunoreactivity of the IgG isotype. Grau et al. [190] further extended their studies by a detailed clinical and biochemical evaluation of infection-associated stroke. In their data on 159 stroke patients without infection and 38 patients with infection, they observed more severe neurological deficits on admission, more frequent cortical infarcts in the middle cerebral artery territory, more frequent cardioembolic strokes, and a smaller prevalence of extracranial artery stenoses.

Heat-shock proteins (hsp's) are a family of proteins of which expression is increased by various stress factors such as high temperature, mechanical stress, and infections. They show highly homologous sequences between different species, from bacteria to man. The hsp65 family has been linked with certain autoimmune diseases because they may evoke autoimmune responses by cross-reaction between hsp's and the host [191]. Arteriosclerosis can be induced in normocholesterolemic rabbits by immunization with hsp65, which is expressed in high concentrations in human atherosclerotic lesions.

Anti-hsp65 antibodies are associated with carotid atherosclerosis in humans, independent of the classic risk factors [192]. It has been postulated that atherosclerosis develops primarily as an autoimmune response aggravated by factors such as hypercholesterolemia [192].

The origin of hsp's in individuals with atherosclerosis is not known. It is of interest that periodontal pathogens and *C. pneumoniae* possess hsp:n with high sequence identity with human hsp [193–196]. Antibodies to the 64-kD protein of *A. actinomycetemcomitans* cross-react in turn with 65-kD proteins of *Haemophilus influenzae* and *E. coli* [194]. Thus, hsp-mediated reactions could be one mechanism linking infection and development of atherosclerosis and thrombosis.

As mentioned earlier, an increasing body of evidence has linked inflammation to CHD. Chronic low-grade infections like those with *C. pneumoniae* and *H. pylori* may well be one of the factors behind this inflammation, which can be detected and measured by an accurate measurement of CRP [2]. In a cross-sectional study of 1,484 patients with angina pectoris, Juhan-Vague et al. [197] showed that plasma insulin levels increased independently of other risk factors with age, body mass index, triglyceride levels, plasminogen activator inhibitor-1, and markers of inflammation, such as CRP.

Mendall et al. [198] investigated the factors determining CRP concentration within the so-called normal range in a random sample of 388 men. *C. pneumoniae* or *H. pylori* seropositivity and an infectious symptom (production of phlegm for 3 months) were among the determinants of CRP levels. Patel et al. [199] observed mutually adjusted differences of 0.43 g/L (95% CI, 0.12–0.75) and 0.52 g/L (95% CI, 0.15–0.9) in fibrinogen levels between persons seropositive for *H. pylori* and *C. pneumoniae* and those who were seronegative; these differences corresponded to 65% and 80% of the standard deviation for fibrinogen, respectively. The investigators have also shown that *C. pneumoniae* and *H. pylori* seropositivity are associated with raised fibrinogen and malondialdehyde concentrations and leukocyte counts [42].

# Conclusions

The idea of infection as a risk factor for atherosclerosis and related diseases is an old one, and infection should not be taken as a "competing view" for the classic coronary risk factors. "New" evidence linking infection to cardiovascular diseases has accumulated since the early 1970s and includes several different infectious agents causing quite different infectious processes: persistent or latent infections like those caused by herpesviruses or *C. pneumoniae;* low-grade bacterial infections like dental infections and *H. pylori* gastritis; and acute, strongly symptomatic infections like bacteremic infections and common upper respiratory infections.

The mechanisms of action probably also vary, ranging from subtle, cytokine-mediated alterations in blood vessel endothelial function to a marked temporary shift of the blood coagulation mechanism to a thrombogenic direction. Likewise, the consequences of these infections vary in different individuals; a marked "procoagulant state" caused by acute infections probably favors thrombus formation in those individuals whose coronary or cerebral arteries are affected by an atherosclerotic process impairing the endothelial functions. On the other hand, *C. pneumoniae*– linked chronic arterial disease seems to be related to the host's HLA status, a circumstance resembling that occurring in the development of reactive arthritis. Infections may also act in concert with the classic coronary risk factors.

Synergy between infection and cholesterol was evident in the studies by Minick et al. [3] and Span et al. [7]. This may well be the case in humans as well. In their prospective 2-year study of 3,043 patients with angina pectoris, Thompson et al. [200] showed that the cholesterol-related relative risk of coronary events did not increase at all in those post-AMI patients who belonged to the lowest CRP tertile, while the risk increased exponentially in individuals in the highest tertile.

Some of the hypotheses generated by the results described earlier have already received support from preliminary results of studies with humans. Gurfinkel et al. reported that the combination of positive *C. pneumoniae* serology and elevated CRP level predicted extremely well future ischemic events in patients with unstable angina, suggesting that the microbe may really be behind the inflammatory reaction observed in these patients [85]. Holme et al. reported a high correlation between *H. pylori*, coronary atherosclerosis, and antibodies to mycobacterial hsp65, again suggesting that it is the infection that causes the autoimmune reaction observed in individuals with atherosclerosis [201].

All in all, as yet the data linking infections to CHD do not prove causality. However, the findings obtained do indicate an urgent need for further studies in this field.

# Addendum

Since preparation of this article, reports of two randomized, double-blind studies showing that antibiotic treatment reduces ischemic events have been published [202, 203]. Although the studies were relatively small, they strongly support a causative role for infections in the pathogenesis of CHD.

#### References

- Osier W. Diseases of the arteries. In: Osier W, ed. Modern medicine: its theory and practice. Philadelphia: Lea & Febiger, 1908:426–47.
- Haverkate F, Thompson S, Pyke SDM, Gallimore JR, Pepys MB, for the European Concerted Action on Thrombosis and Disabilities Angina Pectoris Study Group. Production of C-reactive protein and risk of coronary events in stable and unstable angina. Lancet 1997;349:462–6.
- Minick CR, Fabricant CG, Fabricant J, Litrenta MM. Atheroarteriosclerosis induced by infection with a herpesvirus. Am J Pathol 1979; 96:673–706.
- Fabricant CG, Hajjar DP, Minick CR, Fabricant J. Herpesvirus infection enhances cholesterol and cholesteryl ester accumulation in cultured arterial smooth muscle cells. Am J Pathol 1981;105:176–84.
- Hajjar DP, Pomerantz KB, Falcone DJ, Weksler BB, Grant AJ. Herpes simplex virus infection in human arterial cells: implications in arteriosclerosis. J Clin Invest 1987;80:1317–21.
- Fabricant CG, Fabricant J, Minick CR, Litrenta MM. Herpesvirusinduced atherosclerosis in chickens. Fed Proc 1983;42:2476–9.
- Span AHM, Grauls G, Bosman F, van Boven CPA, Bruggeman CA. Cytomegalovirus infection induces vascular injury in the rat. Atherosclerosis 1992;93:41–52.
- Gyorkey F, Melnick JL, Guinn GA, Gyorkey P, DeBakey ME. Herpesviridae in the endothelial and smooth muscle cells of the proximal aorta in arteriosclerotic patients. Exp Mol Pathol 1984;40:328–39.
- Benditt EP, Barrett T, McDougall JK. Viruses in the etiology of atherosclerosis. Proc Natl Acad Sci 1983;80:6386–9.
- Yamashiroya HM, Ghosh L, Yang R, Robertson AL Jr. Herpesviridae in the coronary arteries and aorta of young trauma victims. Am J Pathol 1988;130:71–9.
- Melnick JL, Petrie BL, Dreesman GR, Burek J, McCollum CH, DeBakey ME. Cytomegalovirus antigen within human arterial smooth muscle cells. Lancet 1983;2:644–7.
- Raza-Ahmad A, Klassen GA, Murphy DA, et al. Evidence of type 2 herpes simplex infection in human coronary arteries at the time of coronary artery bypass surgery. Can J Cardiol **1995**;11:1025–9.
- Hendrix MGR, Salimans MMM, van Boven CPA, Bruggeman CA. High prevalence of latently present cytomegalovirus in arterial walls of patients suffering from grade III atherosclerosis. Am J Pathol 1990;136:23–8.
- Hendrix MGR, Daemen M, Bruggeman CA. Cytomegalovirus nucleic acid distribution within the human vascular tree. Am J Pathol 1991; 138:563-7.
- Melnick JL, Hu C, Burek J, Adam E, DeBakey ME. Cytomegalovirus DNA in arterial walls of patients with atherosclerosis. J Med Virol 1994;42:170–4.
- Adam E, Melnick JL, Probtsfield JL, et al. High levels of cytomegalovirus antibody in patients requiring vascular surgery for atherosclerosis. Lancet 1987;2:291–3.
- Sorlie PD, Adam E, Melnick SL, et al. Cytomegalovirus/herpesvirus and carotid atherosclerosis: the ARIC study. J Med Virol 1994;42:33–7.
- Epstein SE, Speir E, Zhou YF, Guetta E, Leon M, Finkel T. The role of infection in restenosis and atherosclerosis: focus on cytomegalovirus. Lancet 1996; 348(suppl 1):S13–7.
- Speir E, Modali R, Huang ES, et al. Potential role of human cytomegalovirus and p53 interaction in coronary restenosis. Science 1994;265: 391-4.
- Zhou YF, Leon MB, Waclawiw MA, et al. Association between prior cytomegalovirus infection and the risk of restenosis after coronary atherectomy. N Engl J Med 1996;335:624–30.
- Uys CJ, Rose AG. Pathologic findings in long-term cardiac transplants. Arch Pathol Lab Med 1984;108:112-6.
- Demetris AJ, Zerbe T, Banner B. Morphology of solid organ allograft arteriopathy: identification of proliferating intimal cell populations. Transplant Proc 1989;21:3667–9.

- Wu T-C, Hruban RH, Ambinder RF, et al. Demonstration of cytomegalovirus nucleic acids in the coronary arteries of transplanted hearts. Am J Pathol 1992;140:739–47.
- Jäkel KT, Löning T. Herpes virus infections, acute rejection, and transplant arteriosclerosis in human cardiac allografts. Transplant Proc 1993;25:2029–30.
- McDonald K, Rector TS, Braunlin EA, Kubo SH, Olivari MT. Association of coronary artery disease in cardiac transplant recipients with cytomegalovirus infection. Am J Cardiol 1989;64:359–62.
- Grattan MT, Moreno-Cabral CE, Starnes VA, Oyer PE, Stinson EB, Shumway NE. Cytomegalovirus infection is associated with cardiac allograft rejection and atherosclerosis. JAMA 1989;261:3561–6.
- Koskinen PK, Nieminen MS, Krogerius LA, et al. Cytomegalovirus infection and accelerated cardiac allograft vasculopathy in human cardiac allografts. J Heart Lung Transplant 1993;12:724–9.
- Radovancevic B, Pointdexter S, Birovljev S, et al. Risk factors for development of accelerated coronary artery disease in cardiac transplant recipients. Eur J Cardiothorac Surg 1990;4:309–12.
- Sharples LD, Caine N, Mullins P, et al. Risk factor analysis for the major hazards following heart transplantation—rejection, infection, and coronary occlusive disease. Transplantation 1991; 52:244–52.
- Gulizia JM, Kandolf R, Kendall TJ, et al. Infrequency of cytomegalovirus genome in coronary arteriopathy of human heart allografts. Am J Pathol 1995; 147:461–75.
- Paavonen T, Mennender A, Leutenschlager I, Mattila S, Häyry P. Endothelialitis and accelerated arteriosclerosis in human heart transplant coronaries. J Heart Lung Transplant 1993;12:117–22.
- Paton P, Tabib A, Loire R, Tete R. Coronary artery lesions and human immunodeficiency virus infection. Res Virol 1993;144:225-31.
- Kaprio J, Norio R, Pesonen E, Sarna S. Intimal thickening of the coronary arteries in infants in relation to family history of coronary artery disease. Circulation 1993;87:1960–8.
- Minkowski WL. The coronary arteries of infants. Am J Med Sci 1947; 214:623–9.
- Pesonen E, Siitonen O. Acute myocardial infarction precipitated by infectious disease [letter]. Am Heart J 1981;101:512–3.
- Suzuki A, Kamiya T, Arakaki Y, Kinoshita Y, Kimura K. Fate of coronary arterial aneurysms in Kawasaki disease. Am J Cardiol 1994;74: 822–4.
- Kato H, Akagi T, Sugimura T, et al. Kawasaki disease. Coronary Artery Disease 1995;6:194–206.
- Saikku P, Leinonen M, Mattila K, et al. Serological evidence of an association of a novel chlamydia, TWAR, with chronic coronary heart disease and acute myocardial infarction. Lancet 1988;2:983-6.
- Thom DH, Wang S-P, Grayston JT, et al. *Chlamydia pneumoniae* strain TWAR antibody and angiographically demonstrated coronary artery disease. Arterioscler Thromb **1991**;11:547–51.
- Thom DH, Grayston JT, Siscovick DS, Wang S-P, Weiss NS, Daling JR. Association of prior infection with *Chlamydia pneumoniae* and angiographically demonstrated coronary artery disease. JAMA 1992; 268:68–72.
- Melnick SL, Shahar E, Folsom AR, et al. Past infection by *Chlamydia pneumoniae* strain TWAR and asymptomatic carotid atherosclerosis. Am J Med **1993**;95:499–504.
- Patel P, Mendall MA, Carrington D, et al. Association of *Helicobacter* pylori and *Chlamydia pneumoniae* infections with coronary heart disease and cardiovascular risk factors. BMJ 1995;311:711-4.
- 43. Dahlén GH, Boman J, Birgander LS, Lindblom B. Lp(a) lipoprotein, IgG, IgA and IgM antibodies to *Chlamydia pneumoniae* and HLA class II genotype in early coronary artery disease. Atherosclerosis 1995;114:165-74.
- Linnanmäki E, Leinonen M, Mattila K, Nieminen MS, Valtonen VV, Saikku P. Chlamydia pneumoniae-specific circulating immune com-

plexes in patients with chronic coronary heart disease. Circulation **1993**;87:1130-4.

- Shor A, Kuo CC, Patton DL. Detection of *Chlamydia pneumoniae* in coronary arterial fatty streaks and atheromatous plaques. S Afr Med J 1992;82:158–61.
- Kuo C-C, Shor A, Campbell LA, Fukushi H, Patton DL, Grayston JT. Demonstration of *Chlamydia pneumoniae* in atherosclerotic lesions of coronary arteries. J Infect Dis **1993**;167:841–9.
- Kuo C-C, Gown AM, Benditt EP, Grayston JT. Detection of *Chlamydia* pneumoniae in aortic lesions of atherosclerosis by immunocytochemical stain. Arterioseler Thromb **1993**;13:1501–4.
- Campbell LA, O'Brien ER, Cappuccio AL, et al. Detection of *Chlamydia* pneumoniae TWAR in human coronary atherectomy tissues. J Infect Dis **1995**; 172:585–8.
- Grayston JT, Kuo C-C, Coulson AS, et al. *Chlamydia pneumoniae* (TWAR) in atherosclerosis of the carotid artery. Circulation **1995**;92: 3397–400.
- Kuo C-C, Grayston JT, Campbell LA, Goo YA, Wissler RW, Benditt EP. *Chlamydia pneumoniae* (TWAR) in coronary arteries of young adults (15–34 years old). Proc Natl Acad Sci **1995**;92:6911–4.
- Muhlestein J, Hammond EH, Carlquist JF, et al. Increased incidence of *Chlamydia* species within the coronary arteries of patients with symptomatic atherosclerotic versus other forms of cardiovascular disease. J Am Coll Cardiol **1996**;27:1555–61.
- Weiss SM, Roblin PM, Gaydos CA, et al. Failure to detect *Chlamydia* pneumoniae in coronary atheromas of patients undergoing atherectomy. J Infect Dis **1996**;173:957–62.
- Blasi F, Denti F, Erba M, et al. Detection of *Chlamydia pneumoniae* but not *Helicobacter pylori* in atherosclerotic plaques of aortic aneurysms. J Clin Microbiol **1996**; 34:2766–9.
- Juvonen J, Juvonen T, Laurila A, et al. Demonstration of *Chlamydia* pneumoniae in the walls of abdominal aortic aneurysms. J Vasc Surg 1997;25:499–505.
- Puolakkainen M, Kuo C-C, Shor A, Wang S-P, Grayston JT, Campbell LA. Serological response to *Chlamydia pneumoniae* in adults with coronary arterial fatty streaks and fibrolipid plaques. J Clin Microbiol 1993;31:2212–4.
- Ramirez JA, *Chlamydia pneumoniae*/Atherosclerosis Study Group. Isolation of *Chlamydia pneumoniae* from the coronary artery of a patient with coronary atherosclerosis. Ann Intern Med **1996**;125:979–82.
- Cook PJ, Lip GH, Zafris J, Honeyborne D, Wise R, Beevers DG. Is *Chlamydia pneumoniae* infection associated with hypertension? [abstract]. J Hypertens 1995;13:1495–6.
- Loesche WJ. Role of *Streptococcus mutans* in human dental decay. Microbiol Rev 1986;50:353–80.
- Slots J, Listgarten MA. Bacteroides gingivalis, Bacteroides intermedius and Actinobacillus actinomycetemcomitans in human periodontal diseases. J Clin Periodontol 1988;15:85–93.
- 60. Williams RC. Periodontal disease. N Engl J Med 1990; 322:373-82.
- Burt BA. Risk markers for oral disease. In: Johnson NW, ed. Periodontal diseases. Vol. 3. Cambridge: Cambridge University Press, 1991:9–26.
- Vehkalahti M, Paunio K. Kiinnityskudosten kunto [in Finnish]. Kansaneläkelaitoksen julkaisuja 1991; 34:161–81.
- Mattila KJ, Valle MS, Nieminen MS, Valtonen VV, Hietaniemi KL. Dental infections and coronary atherosclerosis. Atherosclerosis 1993; 103:205–11.
- Adams MR, Nakagomi A, Keech A, et al. Carotid intima-media thickness is only weakly correlated with the extent and severity of coronary artery disease. Circulation 1995;92:2127–34.
- Desa'neto A, Bullington JD, Bullington RH, Desser KB, Benchimol A. Coxsackie B5 heart disease: demonstration of inferolateral wall myocardial necrosis. Am J Med 1980;68:295–8.
- Kron J, Lucas L, Lee TD, McAnulty J. Myocardial infarction following an acute viral illness. Arch Intern Med 1983;143:1466–7.

- Miklozek CL, Crumpacker CS, Royal HD, Come PC, Sullivan JL, Abelmann WH. Myocarditis presenting as acute myocardial infarction. Am Heart J 1988;115:768–76.
- Nicholls AC, Thomas M. Coxsackie virus infection in acute myocardial infarction. Lancet 1977;1:883–4.
- Wood SF, Rogen AS, Bell EJ, Grist NR. Role of coxsackie B viruses in myocardial infarction. Br Heart J 1978;40:523-5.
- Griffiths PD, Hannington G, Booth JC. Coxsackie B virus infections and myocardial infarction: results from a prospective epidemiologically controlled study. Lancet **1980**; 1:1387–9.
- Pönkä A, Jalanko H, Pönkä T, Stenvik M. Viral and mycoplasmal antibodies in patients with myocardial infarction. Ann Clin Res 1981;13: 429–32.
- 72. Lau RCH. Coxsackie B virus infection in acute myocardial infarction and adult heart disease. Med J Aust **1982**;2:520–2.
- O'Neill D, McArthur JD, Kennedy JA, Clements G. Coxsackie B virus infection in coronary care unit patients. J Clin Pathol 1983; 36:658–61.
- Nikoskelainen J, Kalliomäki JL, Lapinleimu K, Stenvik M, Halonen PE. Coxsackie B virus antibodies in myocardial infarction. Acta Med Scand 1983;214:29–32.
- Mattila KJ. Viral and bacterial infections in patients with acute myocardial infarction. J Intern Med 1989;225:293-6.
- Kol A, Sperti G, Shani J, et al. Cytomegalovirus replication is not a cause of instability in unstable angina. Circulation 1995;91:1910-3.
- Spodick DH, Flessas AP, Johnson MM. Association of acute respiratory symptoms with onset of acute myocardial infarction: prospective investigation of 150 consecutive patients and matched control patients. Am J Cardiol 1984;53:481–2.
- Bainton D, Jones GR, Hole D. Influenza and ischaemic heart disease a possible trigger for acute myocardial infarction? Int J Epidemiol 1978; 7:231–9.
- Woodhouse PR, Khaw KT, Plummer M, Foley A, Meade TW. Seasonal variation of plasma fibrinogen and factor VII activity in the elderly: winter infections and death from cardiovascular disease. Lancet 1994; 343:435–9.
- Jousilahti P, Vartiainen E, Tuomilehto J, Puska P. Symptoms of chronic bronchitis and the risk of coronary disease. Lancet 1996;348:567–72.
- Saikku P, Leinonen M, Mattila K, et al. Serological evidence of an association of a novel chlamydia, TWAR, with chronic coronary heart disease and acute myocardial infarction. Lancet 1988;2:983-6.
- Leinonen M, Linnanmäki E, Mattila K, et al. Circulating immune complexes containing chlamydial lipopolysaccharide in acute myocardial infarction. Microbiol Pathogenesis 1990;9:67–73.
- Cook PJ, Honeybourne D, Lip GH, Beevers DG, Wise R. *Chlamydia pneumoniae* and acute arterial thrombotic disease [letter]. Circulation 1995;92:3148–9.
- Saikku P, Leinonen M, Tenkanen L, et al. Chronic *Chlamydia pneumoniae* infection as a risk factor for coronary heart disease in the Helsinki Heart Study. Ann Intern Med **1992**;116:273–8.
- Gurfinkel E, Duronto M, Gerda M. Patients with unstable angina related with C reactive protein and *Chlamydia pneumoniae* infection: clinical outcome [abstract P3134]. XVIIIth Congress of the European Society of Cardiology, Birmingham, United Kingdom. Eur Heart J **1996**; 17(suppl):578.
- Miettinen H, Lehto S, Saikku P, et al. Association of *Chlamydia pneu-moniae* and acute coronary heart disease events in non-insulin dependent diabetic and non-diabetic subjects in Finland. Eur Heart J **1996**; 17:682–8.
- Hahn DL, Golubjatnikov R. Smoking is a potential confounder of the *Chlamydia pneumoniae*-coronary artery disease association. Arte-rioscler Thromb 1992;12:945–7.
- Karvonen M, Tuomilehto J, Pitkäniemi J, Naukkarinen A, Saikku P. Importance of smoking for *Chlamydia pneumoniae* seropositivity. Int J Epidemiol **1994**;23:1315–21.

- Karvonen M, Tuomilehto J, Pitkäniemi J, Naukkarinen A, Saikku P. *Chlamydia pneumoniae* IgG antibody prevalence in south-western and eastern Finland in 1982 and 1987. Int J Epidemiol 1994;23:176–84.
- Mattila KJ, Nieminen MS, Valtonen VV, et al. Association between dental health and acute myocardial infarction. BMJ 1989;298:779–81.
- Paunio K, Impivaara O, Tiekso J, Mäki J. Missing teeth and ischaemic heart disease in men aged 45–64 years. Eur Heart J 1993;14(suppl K):54–6.
- DeStefano F, Anda RF, Kahn HS, Williamson DF, Russell CM. Dental disease and risk of coronary heart disease and mortality. BMJ 1993; 306:688–691.
- Mattila KJ, Valtonen VV, Nieminen M, Huttunen JK. Dental infection and the risk of new coronary events: prospective study of patients with documented coronary heart disease. Clin Infect Dis 1995;20:588–92.
- Martîn-de-Argila C, Boixeda D, Cantón R, Gisbert JP, Fuertes A. High seroprevalence of *Helicobacter pylori* infection in coronary heart disease [letter]. Lancet 1995;346:310.
- Morgando A, Sanseverino P, Perotto C, Molino F, Gai V, Ponzetto A. *Helicobacter pylori* seropositivity in myocardial infarction [letter]. Lancet 1995;345:1380.
- Murray LJ, Bamford KB, O'Reilly DPJ, McCrum EE, Evans AE. *Helico-bacter pylori* infection: relation with cardiovascular risk factors, ischaemic heart disease, and social class. Br Heart J **1995**;74:497–501.
- Sandifer QD, Vuilo SU, Crompton G. Association may not be causal [letter]. BMJ 1996;312:251.
- Delaney BC, Hobbs FDR, Holder R. Eradication of the infection on grounds of cardiovascular risk is not supported by current evidence [letter]. BMJ 1996;312:251–2.
- Strandberg TE, Tilvis RS, Vuoristo M, Lindroos M, Kosunen TU. Prospective study of *Helicobacter pylori* seropositivity and cardiovascular diseases in a general elderly population. BMJ **1997**;314:1317–8.
- Lip GYH, Wise R, Beevers G. Association of *Helicobacter pylori* infection with coronary heart disease [letter]. BMJ 1996;312:250–1.
- Mäkelä PH, Mayer H. Enterobacterial common antigen. Bacteriol Rev 1976;40:591-632.
- Valtonen V, Kuikka A, Syrjänen J. Thrombo-embolic complications in bacteraemic infections. Eur Heart J 1993; 14(suppl K):20–3.
- Dowd AB, Grace R, Rees WDW. Cerebral infarction associated with *Mycoplasma pneumoniae* infection [letter]. Lancet 1987;2:567.
- Syrjänen J. Central nervous system complications in patients with bacteremia. Scand J Infect Dis 1989;21:285–96.
- Uza G. Is urinary tract infection a risk factor for ischemic cerebrovascular disease? Med Hypotheses 1987;23:219–24.
- 106. Syrjänen J, Valtonen V, Iivanainen M, Kaste M, Huttunen JK. Preceding infection as an important risk factor for ischaemic brain infarction in young and middle aged patients. Br Med J 1988;296:1156–60.
- Syrjänen J, Peltola J, Valtonen V, Iivanainen M, Kaste M, Huttunen JK. Dental infections in association with cerebral infarction in young and middle-aged men. J Intern Med **1989**;225:179–84.
- Grau AJ, Buggle F, Heindl S, et al. Recent infection as a risk factor for cerebrovascular ischemia. Stroke 1995;26:373–9.
- Wimmer ML, Sandmann-Strupp R, Saikku P, Harberl RL. Association of chlamydial infection with cerebrovascular disease. Stroke 1996;27: 2207–10.
- Basha BJ, Sowers JR. Atherosclerosis: an update. Am Heart J 1996;131: 1192–202.
- 111. Friedman HM, Marcarak EJ, MacGregor RR, Wolfe J, Kefalides NA. Virus infection of endothelial cells. J Infect Dis 1981;143:266-73.
- MacGregor RR, Friedman HM, Macarak EJ, Kefalides NA. Virus infection of endothelial cells increases granulocyte adherence. J Clin Invest 1980;65:1469–77.
- 113. Sacks T, Moldow CF, Craddock PR, Bowers TK, Jacob HS. Oxygen radicals mediate endothelial cell damage by complement-stimulated

granulocytes: an in vitro model of immune vascular damage. J Clin Invest **1978**;61:1161-7.

- Harlan JM, Killen PD, Harker LA, Striker GE. Neutrophil-mediated endothelial injury in vitro: mechanisms of cell detachment. J Clin Invest 1981;68:1394–403.
- Harlan JM. Neutrophil-mediated vascular injury. Acta Med Scand Suppl 1987;715:123–9.
- 116. Tannenbaum SH, Finko R, Cines DB. Antibody and immune complexes induce tissue factor production by human endothelial cells. J Immunol 1986; 137:1532–7.
- 117. Cines DB, Lyss AP, Bina M, Corkey R, Kefalides NA, Friedman HM. Fc and C3 receptors induced by herpes simplex virus on cultured human endothelial cells. J Clin Invest **1982**;69:123–8.
- Visser MR, Tracy PB, Vercellotti GM, Goodman JL, White JG, Jacob HS. Enhanced thrombin generation and platelet binding on herpes simplex virus infected endothelium. Proc Natl Acad Sci USA 1988; 85:8227–30.
- Bok RA, Jacob HS, Balla J, et al. Herpes simplex virus decreases endothelial cell plasminogen activator inhibitor. Thromb Haemostasis 1993; 69:253-8.
- 120. Key NS, Bach RR, Vercellotti GM, Moldow CF. Herpes simplex virus type I does not require productive infection to induce tissue factor in human umbilical vein endothelial cells. Lab Investigation 1993;68: 645-51.
- 121. Jacob HS, Visser M, Kay NS, Goodman JL, Moldow CF, Vercellotti GM. Herpes virus infection of endothelium: new insights into atherosclerosis. Trans Am Clin Climatol Assoc 1992;103:95–104.
- Tumilowicz JJ, Gawlik ME, Powell BB, Trentin JJ. Replication of cytomegalovirus in human arterial smooth muscle cells. J Virol 1985;56: 839–45.
- 123. Bevilacqua MP, Pober JS, Wheeler ME, Cotran RS, Gimbrone MA Jr. Interleukin-1 activation of vascular endothelium: effects on procoagulant activity and leukocyte adhesion. Am J Pathol 1985; 121:394–403.
- 124. Bevilacqua MP, Pober JS, Wheeler ME, Cotran RS, Gimbrone MA Jr. Interleukin 1 acts on cultured human vascular endothelium to increase the adhesion of polymorphonuclear leukocytes, monocytes, and related leukocyte cell lines. J Clin Invest **1985**; 76:2003–11.
- 125. Bevilacqua MP, Pober JS, Majeau GR, Fiers W, Cotran RS, Gimbrone MA Jr. Recombinant tumor necrosis factor induces procoagulant activity in cultured human vascular endothelium: characterization and comparison with the actions of interleukin 1. Proc Natl Acad Sci USA 1986;83:4533-7.
- Miossee P, Cavender D, Ziff M. Production of interleukin 1 by human endothelial cells. J Immunol 1986;136:2486–91.
- 127. Bussolino F, Breviario F, Tetta C, et al. Interleukin 1 stimulates platelet activating factor production in cultured human endothelial cells. Pharmacological Research Communications 1986; 18(suppl):133–7.
- Colucci M, Balconi G, Lorenzet R, et al. Cultured human endothelial cells generate tissue factor in response to endotoxin. J Clin Invest 1983;71:1893-6.
- Hanss M, Collen D. Secretion of tissue-type plasminogen activator and plasminogen activator inhibitor by cultured human endothelial cells: modulation by thrombin, endotoxin, and histamine. J Lab Clin Med 1987; 109:97–104.
- Colucci M, Paramo JA, Collen D. Generation in plasma of a fast-acting inhibitor of plasminogen activator in response to endotoxin stimulation. J Clin Invest 1985;75:818–24.
- Bevilacqua MP, Schleef RR, Gimbrone MA Jr, Loskutoff DJ. Regulation of the fibrinolytic system of cultured human vascular endothelium by interleukin 1. J Clin Invest **1986**; 78:587–91.
- Schorer AE, Moldow CF, Rick ME. Interleukin 1 or endotoxin increases the release of von Willebrand factor from human endothelial cells. Br J Haematol 1987;67:193-7.

- Quesenberry PJ, Gimbrone MA Jr. Vascular endothelium as a regulator of granulopoiesis: production of colony-stimulating activity by cultured human endothelial cells. Blood 1980; 56:1060-7.
- 134. Kaukoranta-Tolvanen SS, Laitinen K, Saikku P, Leinonen M. Chlamydia pneumoniae multiplies in human endothelial cells in vitro. Microb Pathog 1994;16:313–9.
- Kaukoranta-Tolvanen SS. Pathogenesis of *Chlamydia pneumoniae* infection [thesis]. Helsinki: University of Helsinki, **1996**:1–77.
- Duncan RL Jr, Morrison DC. The fate of *E. coli* lipopolysaccharide after the uptake of *E. coli* by murine macrophages in vitro. J Immunol **1984**; 132:1416–24.
- 137. Assoian RK, Fleurdelys BE, Stevenson HC, et al. Expression and secretion of type β transforming growth factor by activated human macrophages. Proc Natl Acad Sci USA 1987;84:6020–4.
- Glenn KC, Ross R. Human monocyte-derived growth factor(s) for mesenchymal cells: activation of secretion by endotoxin and concanavalin A. Cell 1981;25:603–15.
- DeLustro F, LeRoy EC. Characterization of the release of human monocyte regulators of fibroblast proliferation. J Reticuloendothelial Soc 1982;31:295–305.
- 140. Martinet Y, Bitterman PB, Mornex J-F, Grotendorst GR, Martin GR, Crystal RG. Activated human monocytes express the *c-sis* proto-oncogene and release a mediator showing PDGF-like activity. Nature 1986; 319:158–60.
- 141. Wahl SM, McCartney-Francis N, Hunt DA, Smith PD, Wahl LM, Katona IM. Monocyte interleukin 2 receptor gene expression and interleukin 2 augmentation in microbicidal activity. J Immunol **1987**;139:1342–7.
- Lepe-Zuniga JL, Gery I. Production of intra- and extracellular interleukin-1 (IL-1) by human monocytes. Clin Immunol Immunopathol 1984;31: 222–30.
- 143. Libby P, Ordovas JM, Auger KR, Robbins AH, Birinyi LK, Dinarello CA. Endotoxin and tumor necrosis factor induce interleukin-1 gene expression in adult human vascular endothelial cells. Am J Pathol 1986; 124:179–85.
- 144. Di Corleto PE, Chisolm GM III. Participation of the endothelium in the development of the atherosclerotic plaque. Prog Lipid Res 1986;25: 365-74.
- Pugin J, Ulevitch RJ, Tobias PS. Activation of endothelial cells by endotoxin: direct versus indirect pathways and the role of CD14. Prog Clin Biol Res 1995; 392:369–73.
- Mazzone T, Jensen M, Chait A. Human arterial wall cells secrete factors that are chemotactic for monocytes. Proc Natl Acad Sci USA 1983; 80:5094–7.
- 147. Seatter SC, Li MH, Bubrick MP, West MA. Endotoxin pretreatment of human monocytes alters subsequent endotoxin-triggered release of inflammatory mediators. Shock 1995;3:252–8.
- Schäfer C, Schips I, Landig J, Bode JC, Bode C. Tumor-necrosis-factor and interleukin-6 response of peripheral blood monocytes to low concentrations of lipopolysaccharide in patients with alcoholic liver disease. Z Gastroenterol 1995; 33:503–8.
- 149. MacFarlane CG, Reynolds JJ, Meikle MC. The release of interleukin-1β, tumor necrosis factor-α, and interferon-γ by cultured peripheral blood mononuclear cells from patients with periodontitis. J Periodont Res 1990;25:207–14.
- Ernst E, Hammerschmidt DE, Bagge U, Matrai A, Dormandy JA. Leukocytes and the risk of ischemic diseases. JAMA 1987;257:2318–24.
- Jacob HS, Craddock PR, Hammerschmidt DE, Moldow CF. Complement-induced granulocyte aggregation: an unsuspected mechanism of disease. N Engl J Med 1980; 302:789–94.
- 152. Guthrie LA, McPhail LC, Henson PM, Johnston RB Jr. Priming of neutrophils for enhanced release of oxygen metabolites by bacterial lipopolysaccharide: evidence for increased activity of the superoxide-producing enzyme. J Exp Med **1984**; 160:1656–71.

- 153. Østerud B. Interaction of endotoxins, blood elements and the vessel wall. In: ten Cate JW, Büller HR, Sturk A, Levin J, eds. Bacterial endotoxins: structure, biomedical significance, and detection with the limulus amebocyte lysate test. Progress in clinical and biological research. Vol. 189. New York: Alan R. Liss, **1985**:67–78.
- 154. Fittschen C, Sandhaus RA, Worthen GS, Henson PM. Bacterial lipopolysaccharide enhances chemoattractant-induced elastase secretion by human neutrophils. J Leukocyte Biol **1988**;43:547–56.
- 155. Lewis MS, Whatley RE, Cain P, McIntyre TM, Prescott SM, Zimmerman GA. Hydrogen peroxide stimulates the synthesis of platelet-activating factor by endothelium and induces endothelial cell-dependent neutrophil adhesion. J Clin Invest 1988;82:2045–55.
- 156. Worthen GS, Seccombe JF, Clay KL, Guthrie LA, Johnston RB Jr. The priming of neutrophils by lipopolysaccharide for production of intracellular platelet-activating factor: potential role in mediation of enhanced superoxide secretion. J Immunol **1988**; 140:3553–9.
- Vercellotti GM, Yin HQ, Gustafson KS, Nelson RD, Jacob HS. Plateletactivating factor primes neutrophil responses to agonists: role in promoting neutrophil-mediated endothelial damage. Blood 1988;71:1100–7.
- Kapp A, Luger TA, Maly FE, Schöpf E. Granulocyte-activating mediators (GRAM): I. Generation by lipopolysaccharide-stimulated mononuclear cells. J Invest Dermatol 1986;86:523–8.
- 159. Tzeng DY, Deuel TF, Huang JS, Senior RM, Boxer LA, Bachner RL. Platelet-derived growth factor promotes polymorphonuclear leukocyte activation. Blood **1984**;64:1123–8.
- Ozaki Y, Ōhashi T, Niwa Y. Oxygen radical production by neutrophils from patients with bacterial infection and rheumatoid arthritis. Inflammation 1986; 10:119–30.
- Suffredini AF, Harpel PC, Parrillo JE. Promotion and subsequent inhibition of plasminogen activation after administration of intravenous endotoxin to normal subjects. N Engl J Med 1989;320:1165–72.
- Gustafsson A, Åsman B. Increased release of free oxygen radicals from peripheral neutrophils in adult periodontitis after Fc-receptor stimulation. J Clin Periodontol 1996;23:38–44.
- 163. Kimura S, Yonemura T, Kaya H. Increased oxidative product formation by peripheral blood polymorphonuclear leukocytes in human periodontal diseases. J Periodont Res 1993;28:197–203.
- 164. Kweider M, Lowe GD, Murray GD, Kinane DF, McGowan DA. Dental disease, fibrinogen and white cell count: links with myocardial infarction? Scott Med J 1993; 38:73–4.
- 165. Van Lenten BJ, Fogelman AM, Haberland ME, Edwards PA. The role of lipoproteins and receptor-mediated endocytosis in the transport of bacterial lipopolysaccharide. Proc Natl Acad Sci USA 1986;83: 2704–8.
- 166. Wurfel MM, Hailman E, Wright SD. Soluble CD14 acts as a shuttle in the neutralization of lipopolysaccharide (LPS) by LPS-binding protein and reconstituted high density lipoprotein. J Exp Med 1995;181: 1743–54.
- 167. Liao W, Florén C-H. Hyperlipidemic response to endotoxin—a part of the host-defence mechanism. Scand J Infect Dis 1993;25:675–82.
- Feingold KR, Funk JL, Moser AH, Shigenaga JK, Rapp JH, Grunfeld C. Role of circulating lipoproteins in protection from endotoxin toxicity. Infect Immun 1995;63:2041–6.
- Morel DW, Di Corleto PE, Chisolm GM. Modulation of endotoxininduced endothelial cell toxicity by low-density lipoprotein. Lab Invest 1986; 55:419–26.
- 170. Brand K, Banka CL, Mackman N, Terkeltaub RA, Fan S-T, Curtiss LK. Oxidized LDL enhances lipopolysaccharide-induced tissue factor expression in human adherent monocytes. Arterioscler Thromb 1994; 14:790–7.
- 171. Cathcart MK, Chisolm GM III, McNally AK, Morel DW. Oxidative modification of low density lipoprotein (LDL) by activated human monocytes and the cell lines U937 and HL60. In Vitro Cell Develop Biol **1988**;24:1001–8.

- Lopes-Virella MF, Klein RL, Stevenson HC. Low density lipoprotein metabolism in human macrophages stimulated with microbial or microbial-related products. Arteriosclerosis 1987;7:176–84.
- 173. Wiernik A, Carlson LA, Jarstrand C. High-density lipoproteins inhibit the bacterial lipopolysaccharide-mediated increase in oxidative metabolism and lysozyme release by neutrophilic granulocytes in vitro. J Clin Lab Immunol **1986**;21:131–5.
- 174. Sammalkorpi K, Valtonen V, Kerttula Y, Nikkilä E, Taskinen M-R. Changes in serum lipoprotein pattern induced by acute infections. Metabolism 1988;37:859–65.
- Clawson CC, Rao GHR, White JG. Platelet interaction with bacteria. IV. Stimulation of the release reaction. Am J Pathol 1975;81:411–20.
- Kurpiewski GE, Forrester LJ, Campbell BJ, Barrett JT. Platelet aggregation by *Streptococcus pyogenes*. Infect Immun 1983; 39:704–8.
- Forrester LJ, Campbell BJ, Berg JN, Barrett JT. Aggregation of platelets by *Fusobacterium necrophorum*. J Clin Microbiol 1985;22:245–9.
- Herzberg MC, Brintzenhofe KL, Clawson CC. Aggregation of human platelets and adhesion of *Streptococcus sanguis*. Infect Immun 1983; 39:1457–69.
- Miragliotta G, Del Prete R, Mosca A. *Helicobacter pylori* infection and coronary heart disease [letter]. Lancet 1994;344:751.
- Schwartz BS, Monroe MC. Human platelet aggregation is initiated by peripheral blood mononuclear cells exposed to bacterial lipopolysaccharide in vitro. J Clin Invest 1986; 78:1136–41.
- 181. Tsao BP, Fair DS, Curtiss LK, Edgington TS. Monocytes can be induced by lipopolysaccharide-triggered T-lymphocytes to express functional factor VII/VIIa protease activity. J Exp Med 1984;159:1042–57.
- 182. Schwartz BS, Monroe MC, Levin EG. Increased release of plasminogen activator inhibitor type 2 accompanies the human mononuclear cell tissue factor response to lipopolysaccharide. Blood 1988;71: 734-41.
- Bjornson HS. Activation of Hageman factor by lipopolysaccharides of Bacteroides fragilis, Bacteroides vulgatus, and Fusobacterium mortiferum. Rev Infect Dis 1984;6(suppl 1):S30–3.
- 184. Zahavi J. The role of platelets in myocardial infarction, ischemic heart disease, cerebrovascular disease, thromboembolic disorders, and acute idiopathic pericarditis. Thromb Haemost 1977; 38:1073–84.
- Rasi V, Ikkala E, Valtonen V. Plasma β-thromboglobulin in severe infection. Thromb Res 1982;26:267–74.
- Richardson GN, Matthews KB, Cruickshank JK, Geddes AM, Stuart J. Coagulation activation and hyperviscosity in infection. Br J Haematol 1979;42:469–80.
- 187. Mattila K, Rasi V, Nieminen M, et al. Von Willebrand factor antigen and dental infections. Thrombosis Research 1989;56:325–9.
- Olsanska-Seidlová A, Skarlandt P, Mikulecky M, Seymour G. Some immunological findings in adult periodontitis. Aust Dent J 1989;34:417–20.
- Ameriso SF, Wong VLY, Quismorio FP Jr, Fisher M. Immunohematologic characteristics of infection-associated cerebral infarction. Stroke 1991;22:1004–9.
- Grau AJ, Buggle F, Steichen-Wiehn C, et al. Clinical and biochemical analysis in infection-associated stroke. Stroke 1995;26:1520-6.
- Kaufmann SHE. Heat shock proteins and immune response. Immunol Today 1990; 11:129–36.
- Xu Q, Dietrich H, Steiner HJ, et al. Induction of arteriosclerosis in normocholesterolemic rabbits by immunization with heat shock protein 65. Arterioscler Thromb **1992**; 12:789–99.
- 193. Vayssier C, Mayrand D, Grenier D. Detection of stress proteins in *Porphyromonas gingivalis* and other oral bacteria by western immunoblotting analysis. FEMS Microbiol Lett **1994**;121:303–8.
- 194. Nakano Y, Inai Y, Yamashita Y, et al. Molecular and immunological characterization of a 64-kDa protein of *Actinobacillus actinomycetemcomitans*. Oral Microbiol Immunol **1995**;10:151–9.
- 195. Ando T, Kato T, Ishihara K, Ogiuchi H, Okuda K. Heat shock proteins

in the human periodontal disease process. Microbiol Immunol **1995**; 39:321–7.

- Kikuta LC, Puolakkainen M, Kuo C-C, Campbell LA. Isolation and sequence analysis of the *Chlamydia pneumoniae* GroE operon. Infect Immun 1991; 59:4665–9.
- 197. Juhan-Vague I, Thompson SG, Jespersen TH, and the ECAT Angina Pectoris Study Group. Involvement of the hemostatic system in the insulin resistance syndrome: a study of 1500 patients with angina pectoris. Arterioscler Thromb **1993**; 13:1865–73.
- 198. Mendall MA, Patel P, Ballam L, Strachan D, Northfield TC. C reactive protein and its relation to cardiovascular risk factors: a population based cross sectional study. BMJ 1996;312:1061–5.
- Patel P, Carrington D, Strachan DP, et al. Fibrinogen: a link between chronic infection and coronary heart disease [letter]. Lancet 1994;343: 1634–5.



- 200. Thompson SG, Kienast J, Pyke SDM, Haverkate F, van de Loo JCW. Hemostatic factors and the risk of myocardial infarction or sudden death in patients with angina pectoris. N Engl J Med 1995; 332: 635–41.
- Birnie Holme E, McKay IC, Hood S, McKoll KEL, Hills WS. Correlation between antibodies to heat shock protein 65 and coronary atherosclerosis: possible role of *H. pylori* infection [abstract 1345]. XVIIIth Congress of the Society of Cardiology (Birmingham, UK). **1996**:231.
- 202. Gurfinkel E, Bozovich G, Daroca A, Beck E, Mautner B. Randomised trial of roxithromycin in non-Q-wave coronary syndromes: ROXIS Pilot Study. ROXIS Study Group. Lancet **1997**;350:404–7.
- 203. Gupta S, Leatham EW, Carrington D, Mendall MA, Kaski JC, Camm AJ. Elevated *Chlamydia pneumoniae* antibodies, cardiovascular events, and azithromycin in male survivors of myocardial infarction. Circulation **1997**;96:404–7.

This stamp was issued by the Dominican Republic on 2 June 1979 to publicize the Dominican Cardiology Institute. It illustrates a thrombosed atherosclerotic artery overlying a heart with an occluded coronary artery and an acute myocardial infarct. (From the medical philately collection of Dr. J. N. Shanberge, University of Michigan.)